Genetic Architecture of Subcortical Brain Structures in 38,851 Individuals

Claudia L Satizabal*[1-4], Hieab HH Adams*[5-7], Derrek P Hibar*[8], Charles C White*[9,10], Maria J Knol[5], Jason L Stein[8,11,12], Markus Scholz[13,14], Muralidharan Sargurupremraj[15], Neda Jahanshad[8], Gennady V Roshchupkin[5,6,16], Albert V Smith[17-19], Joshua C Bis[20], Xueqiu Jian[21], Michelle Luciano[22], Edith Hofer[23,24], Alexander Teumer[25], Sven J van der Lee[5], Jingyun Yang[26,27], Lisa R Yanek[28], Tom V Lee[29], Shuo Li[30], Yanhui Hu[31], Jia Yu Koh[32], John D Eicher[33], Sylvane Desrivières[34], Alejandro Arias-Vasquez[35-38], Ganesh Chauhan[39,40], Lavinia Athanasiu[41,42], Miguel E Rentería[43], Sungeun Kim[44-46], David Hoehn[47], Nicola J Armstrong[48], Qiang Chen[49], Avram J Holmes[50,51], Anouk den Braber[52-55], Iwona Kloszewska[56], Micael Andersson[57,58], Thomas Espeseth[59,60], Oliver Grimm[61], Lucija Abramovic[62], Saud Alhusaini[63,64], Yuri Milaneschi[65], Martina Papmeyer[66,67], Tomas Axelsson[68], Stefan Ehrlich[51,69,70], Roberto Roiz-Santiañez[71-73], Bernd Kraemer[74], Asta K Håberg[75,76], Hannah J Jones[77-79], G Bruce Pike[80,81], Dan J Stein[82,83], Allison Stevens[70], Janita Bralten[36,38], Meike W Vernooij[5,6], Tamara B Harris[84], Irina Filippi[85], A Veronica Witte[86,87], Tulio Guadalupe[88,89], Katharina Wittfeld[90,91], Thomas H Mosley[92], James T Becker[93-95], Nhat Trung Doan[96], Saskia P Hagenaars[22], Yasaman Saba[97], Gabriel Cuellar-Partida[98], Najaf Amin[5], Saima Hilal[99,100], Kwangsik Nho[44-46], Nazanin Mirza-Schreiber[47,101], Konstantinos Arfanakis[26,102,103], Diane M Becker[28], David Ames[104,105], Aaron L Goldman[49], Phil H Lee[51,106-109], Dorret I Boomsma[52-54,110], Simon Lovestone[111,112], Sudheer Giddaluru[113,114], Stephanie Le Hellard[113,114], Manuel Mattheisen[115-119], Marc M Bohlken[62], Dalia Kasperaviciute[120,121], Lianne Schmaal[122,123], Stephen M Lawrie[66], Ingrid Agartz[96,118,124], Esther Walton[69,125], Diana Tordesillas-

Gutierrez[73,126], Gareth E Davies[127], Jean Shin[128], Jonathan C Ipser[82], Louis N Vinke[129], Martine Hoogman[36,38], Tianye Jia[34], Ralph Burkhardt[14,130], Marieke Klein[36,38], Fabrice Crivello[131], Deborah Janowitz[90], Owen Carmichael[132], Unn K Haukvik[133,134], Benjamin S Aribisala[135,136], Helena Schmidt[97], Lachlan T Strike[98,137], Ching-Yu Cheng[32,138], Shannon L Risacher[45,46], Benno Pütz[47], Debra A Fleischman[26,27,139], Amelia A Assareh[140], Venkata S Mattay[49,141,142], Randy L Buckner[51,143], Patrizia Mecocci[144], Anders M Dale[145-149], Sven Cichon[150-152], Marco P Boks[62], Mar Matarin[120,153,154], Brenda WJH Penninx[65], Vince D Calhoun[155-157], M Mallar Chakravarty[158,159], Andre Marquand[38,160], Christine Macare[34], Shahrzad Kharabian Masouleh[86,161], Jaap Oosterlaan[162-164], Philippe Amouyel[165-168], Katrin Hegenscheid[169], Jerome I Rotter[170], Andrew J Schork[171,172], David CM Liewald[22], Greig I De Zubicaray[173,174], Tien Yin Wong[32,175], Li Shen[175,176], Philipp G Sämann[47], Henry Brodaty[140,177], Joshua L Roffman[51], Eco JC De Geus[52-54,110], Magda Tsolaki[178], Susanne Erk[179], Kristel R Van Eijk[180], Gianpiero L Cavalleri[181], Nic JA Van der Wee[182,183], Andrew M McIntosh[22,66], Randy L Gollub[51,70,106], Kazima B Bulayeva[184], Manon Bernard[128], Jennifer S Richards[35,38,185], Jayandra J Himali[3,4,30], Markus Loeffler[13,14], Nanda Rommelse[37,38,186], Wolfgang Hoffmann[91,187], Lars T Westlye[59,188], Maria C Valdés Hernández[135,189], Narelle K Hansell[98,137], Theo GM Van Erp[190,191], Christiane Wolf[192], John BJ Kwok[193-195], Bruno Vellas[196,197], Andreas Heinz[198], Loes M Olde Loohuis[199], Norman Delanty[63,200], Beng-Choon Ho[201], Christopher RK Ching[8,202], Elena Shumskaya[36,38,160], Baljeet Singh[203], Albert Hofman[5,204], Dennis Van der Meer[205,206], Georg Homuth[207], Bruce M Psaty[20,208-210], Mark E Bastin[135,189], Grant W Montgomery[211], Tatiana M Foroud[46,212], Simone Reppermund[140,213], Jouke-Jan Hottenga[52-54,110], Andrew Simmons[214-216], Andreas Meyer-Lindenberg[61], Wiepke

Cahn[62], Christopher D Whelan[8,63], Marjolein MJ Van Donkelaar[36,38], Qiong Yang[30], Norbert Hosten[169], Robert C Green[106,217], Anbupalam Thalamuthu[140], Sebastian Mohnke[179], Hilleke E Hulshoff Pol[62], Honghuang Lin[3,218], Clifford R Jack Jr[219], Peter R Schofield[194,220], Thomas W Mühleisen[152,221,222], Pauline Maillard[203], Steven G Potkin[223], Wei Wen[140], Evan Fletcher[203], Arthur W Toga[224], Oliver Gruber[74], Matthew Huentelman[172], George Davey Smith[78], Lenore J Launer[84], Lars Nyberg[57,225,226], Erik G Jönsson[96,118], Benedicto Crespo-Facorro[72,73], Nastassja Koen[82,83], Douglas N Greve[70,227], André G Uitterlinden[5,228], Daniel R Weinberger[49,141,229-231], Vidar M Steen[113,114], Iryna O Fedko[52,53,110], Nynke A Groenewold[82], Wiro J Niessen[6,16,232], Roberto Toro[233], Christophe Tzourio[40], William T Longstreth Jr[209,234], M Kamran Ikram[5,235], Jordan W Smoller[51,106,108,109], Marie-Jose Van Tol[236], Jessika E Sussmann[66], Tomas Paus[237-239], Hervé Lemaître[85], Matthias L Schroeter[14,86,240], Bernard Mazoyer[131], Ole A Andreassen[59,96], Florian Holsboer[47,241], Chantal Depondt[242], Dick J Veltman[65], Jessica A Turner[156,157,243], Zdenka Pausova[128], Gunter Schumann[34], Daan Van Rooij[35,38,185], Srdjan Djurovic[113,244], Ian J Deary[22], Katie L McMahon[173,174], Bertram Müller-Myhsok[47,245,246], Rachel M Brouwer[62], Hilkka Soininen[247,248], Massimo Pandolfo[242], Thomas H Wassink[201], Joshua W Cheung[8], Thomas Wolfers[36,38], Jean-Luc Martinot[249], Marcel P Zwiers[38,160], Matthias Nauck[250,251], Ingrid Melle[59,96], Nicholas G Martin[98], Ryota Kanai[252-254], Eric Westman[255], René S Kahn[62], Sanjay M Sisodiya[120], Tonya White[6,256], Arvin Saremi[8], Hans van Bokhoven[36,38], Han G Brunner[36,38,257,258], Henry Völzke[25,251], Margaret J Wright[137,259], Dennis Van 't Ent[52-54,110], Markus M Nöthen[151,260], Roel A Ophoff[62,199], Jan K Buitelaar[35,38,186], Guillén Fernández[35,38], Perminder S Sachdev[140,261], Marcella Rietschel[61], Neeltje EM Van Haren[62,256], Simon E Fisher[38,89], Alexa S Beiser[3,4,30], Clyde Francks[38,89],

Andrew J Saykin[45,46,212], Karen A Mather[140,194], Nina Romanczuk-Seiferth[198], Catharina A Hartman[185], Anita L DeStefano[3,30], Dirk J Heslenfeld[262], Michael W Weiner[263,264], Henrik Walter[179], Pieter J Hoekstra[185], Paul A Nyquist[28], Barbara Franke[36-38], David A Bennett[26,27], Hans J Grabe[90,91], Andrew D Johnson[33], Christopher Chen[99,100], Cornelia M van Duijn[5,265], Oscar L Lopez[94,95], Myriam Fornage[21,266], Joanna M Wardlaw[22,189,267], Reinhold Schmidt[23], Charles DeCarli[268], Philip L De Jager[269,270], Arno Villringer[86,87], Stéphanie Debette[40], Vilmundur Gudnason[18,19], Sarah E Medland[98]**, Joshua M Shulman[29,271-273]**, Paul M Thompson[8]**, Sudha Seshadri[1,3,4]**, M Arfan Ikram[5,6]**

1. Glenn Biggs Institute for Alzheimer's & Neurodegenerative Diseases, UT Health San Antonio, San Antonio, Texas, USA.
2. Department of Population Health Sciences, UT Health San Antonio, San Antonio, Texas, USA.
3. The Framingham Heart Study, Framingham, Massachusetts, USA.
4. Department of Neurology, Boston University School of Medicine, Boston, Massachusetts, USA.
5. Department of Epidemiology, Erasmus MC, Rotterdam, The Netherlands.
6. Department of Radiology and Nuclear Medicine, Erasmus MC, Rotterdam, The Netherlands.
7. Department of Clinical Genetics, Erasmus MC, Rotterdam, The Netherlands.
8. Imaging Genetics Center, USC Mark and Mary Stevens Neuroimaging and Informatics Institute, Keck School of Medicine, University of Southern California, Los Angeles, USA.
9. Cell Circuits Program, Broad Institute, Cambridge, Massachusetts, USA.
10. Center for Translational & Computational Neuroimmunology, Department of Neurology, Columbia University Medical Center, New York City, New York, USA.

11. Department of Genetics, University of North Carolina (UNC), Chapel Hill, North Carolina, USA.
12. UNC Neuroscience Center, University of North Carolina, Chapel Hill, North Carolina, USA.
13. Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Leipzig, Germany.
14. LIFE - Leipzig Research Center for Civilization Diseases, University of Leipzig, Leipzig, Germany.
15. INSERM U1219, VINTAGE team, University of Bordeaux, Bordeaux, France.
16. Department of Medical Informatics, Erasmus MC, Rotterdam, The Netherlands.
17. Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, USA.
18. Faculty of Medicine, University of Iceland, Iceland.
19. Icelandic Heart Association, Iceland.
20. Cardiovascular Health Research Unit, Department of Medicine, University of Washington, Seattle, Washington, USA.
21. The Brown Foundation Institute of Molecular Medicine, University of Texas Health Science Center at Houston, Houston, Texas, USA.
22. Centre for Cognitive Ageing and Cognitive Epidemiology, Psychology, University of Edinburgh, Edinburgh, UK.
23. Clinical Division of Neurogeriatrics, Department of Neurology, Medical University of Graz, Graz, Austria.
24. Institute for Medical Informatics, Statistics and Documentation, Medical University of Graz, Graz, Austria.
25. Institute for Community Medicine, University Medicine Greifswald, Greifswald, Germany.
26. Rush Alzheimer's Disease Center, Rush University Medical Center, Chicago, Illinois, USA.

27. Department of Neurological Sciences, Rush University Medical Center, Chicago, Illinois, USA.
28. GeneSTAR Research Program, Department of Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.
29. Department of Neurology, Baylor College of Medicine, Houston, Texas, USA.
30. Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA.
31. Department of Genetics, Harvard Medical School, Boston, Massachusetts, USA.
32. Singapore Eye Research Institute, Singapore National Eye Centre, Singapore.
33. National Heart, Lung and Blood Institute's Framingham Heart Study, Division of Intramural Research, Population Sciences Branch, Framingham, Massachusetts, USA.
34. MRC-SGDP Centre, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, UK.
35. Department of Cognitive Neuroscience, Radboud University Medical Center, Nijmegen, The Netherlands.
36. Department of Human Genetics, Radboud University Medical Center, Nijmegen, The Netherlands.
37. Department of Psychiatry, Radboud University Medical Center, Nijmegen, The Netherlands.
38. Donders Institute for Brain, Cognition and Behaviour, Radboud University, Nijmegen, The Netherlands.
39. Centre for Brain Research, Indian Institute of Science, Bangalore, India.
40. Univ. Bordeaux, Inserm, Bordeaux Population Health Research Center, UMR 1219, CHU Bordeaux, Bordeaux, France.
41. CoE NORMENT, Division of Mental Health and Addiction, Oslo University Hospital, Oslo, Norway.

42. CoE NORMENT, Institute of Clinical Medicine, University of Oslo, Oslo, Norway.
43. Department of Genetics and Computational Biology, QIMR Berghofer Medical Research Institute, Brisbane, Queensland, Australia.
44. Center for Computational Biology and Bioinformatics, Indiana University School of Medicine, Indianapolis, Indiana, USA.
45. Center for Neuroimaging, Radiology and Imaging Sciences, Indiana University School of Medicine, Indianapolis, Indiana, USA.
46. Indiana Alzheimer Disease Center, Indiana University School of Medicine, Indianapolis, Indiana, USA.
47. Max Planck Institute of Psychiatry, Munich, Germany.
48. Mathematics and Statistics, Murdoch University, Perth, Western Australia, Australia.
49. Lieber Institute for Brain Development, Baltimore, Maryland, USA.
50. Department of Psychology, Yale University, New Haven, Connecticut, USA.
51. Department of Psychiatry, Massachusetts General Hospital, Boston, Massachusetts, USA.
52. Department of Biological Psychology, Vrije Universiteit Amsterdam, Amsterdam, The Netherlands.
53. Netherlands Twin Register, Vrije Universiteit, Amsterdam, The Netherlands.
54. Amsterdam Neuroscience, Amsterdam, The Netherlands.
55. Alzheimer Center Amsterdam, Department of Neurology, Amsterdam Neuroscience, VU Amsterdam, Amsterdam UMC, Amsterdam, The Netherlands.
56. Medical University of Lodz, Lodz, Poland.
57. Department of Integrative Medical Biology, Umeå University, Umeå, Sweden.
58. Umeå Center for Functional Brain Imaging, Umeå University, Umeå, Sweden.

59. NORMENT - KG Jebsen Centre for Psychosis Research, Division of Mental Health and Addiction, Oslo University Hospital, Oslo, Norway.

60. Department of Psychology, University of Oslo, Oslo, Norway.

61. Central Institute of Mental Health, Medical Faculty Mannheim, University Heidelberg, Mannheim, Germany.

62. Department of Psychiatry, UMC Brain Center, University Medical Center Utrecht, Utrecht University, Utrecht, The Netherlands.

63. The Royal College of Surgeons in Ireland, Dublin, Ireland.

64. Department of Neurology, Yale School of Medicine, New Haven, Connecticut, USA.

65. Department of Psychiatry, Amsterdam Neuroscience, VU University Medical Center, Amsterdam, The Netherlands

66. Division of Psychiatry, Royal Edinburgh Hospital, University of Edinburgh, Edinburgh, UK.

67. Division of Systems Neuroscience of Psychopathology, Translational Research Center, University Hospital of Psychiatry, University of Bern, Bern, Switzerland.

68. Department of Medical Sciences, Molecular Medicine and Science for Life Laboratory, Uppsala University, Uppsala, Sweden.

69. Division of Psychological and Social Medicine and Developmental Neurosciences, Faculty of Medicine, TU Dresden, Dresden, Germany.

70. Martinos Center for Biomedical Imaging, Massachusetts General Hospital, Charlestown, Massachusetts, USA.

71. Department of Psychiatry, University Hospital Marqués de Valdecilla, School of Medicine, University of Cantabria-IDIVAL, Santander, Spain.

72. Department of Medicine, University Hospital Marqués de Valdecilla, School of Medicine, University of Cantabria-IDIVAL, Santander, Spain.

73. CIBERSAM (Centro Investigación Biomédica en Red Salud Mental), Santander, Spain.

74. Section for Experimental Psychopathology and Neuroimaging, Dept of General Psychiatry, Heidelberg University, Heidelberg, Germany.

75. Department of Neuroscience, Faculty of Medicine, Norwegian University of Science and Technology (NTNU), Trondheim, Norway.

76. Department of Radiology, St. Olav's Hospital, Trondheim University Hospital, Trondheim, Norway.

77. Centre for Academic Mental Health, Population Health Sciences, Bristol Medical School, University of Bristol, Bristol, UK.

78. MRC Integrative Epidemiology Unit, Bristol Medical School, University of Bristol, Bristol, UK.

79. NIHR Biomedical Research Centre at the University Hospitals Bristol NHS Foundation Trust and the University of Bristol, Bristol UK.

80. Department of Radiology, University of Calgary, Calgary, Alberta, Canada.

81. Department of Clinical Neurosciences, University of Calgary, Calgary, Alberta, Canada.

82. Department of Psychiatry and Mental Health, University of Cape Town, Cape Town, South Africa.

83. South African Medical Research Council (SAMRC) Unit on Risk & Resilience in Mental Disorders, Cape Town, South Africa.

84. Laboratory of Epidemiology and Population Sciences, National Institute on Aging, Intramural Research Program, National Institutes of Health, Bethesda, Maryland, USA.

85. INSERM UMR 1000 ''Neuroimaging and Psychiatry'', University Paris-Sud, University Paris-Saclay, University Paris Descartes, Paris, France.

86. Department of Neurology, Max Planck Institute for Human Cognitive and Brain Sciences, Leipzig, Germany.

87. Faculty of Medicine, CRC 1052 Obesity Mechanisms, University of Leipzig, Leipzig, Germany.

88. International Max Planck Research School for Language Sciences, Nijmegen, The Netherlands.

89. Language and Genetics Department, Max Planck Institute for Psycholinguistics, Nijmegen, The Netherlands.

90. Department of Psychiatry, University Medicine Greifswald, Greifswald, Germany.

91. German Center for Neurodegenerative Diseases (DZNE), Rostock/Greifswald, Greifswald, Germany.

92. Department of Medicine, University of Mississippi Medical Center, Jackson, Mississippi, USA.

93. Department of Psychology, University of Pittsburgh, Pittsburgh, Pennsylvania, USA.

94. Department of Neurology, University of Pittsburgh, Pittsburgh, Pennsylvania, USA.

95. Department of Psychiatry, University of Pittsburgh, Pittsburgh, Pennsylvania, USA.

96. NORMENT - KG Jebsen Centre for Psychosis Research, Institute of Clinical Medicine, University of Oslo, Oslo, Norway.

97. Institute of Molecular Biology and Biochemistry, Centre for Molecular Medicine, Medical University of Graz, Graz, Austria.

98. QIMR Berghofer Medical Research Institute, Brisbane, Queensland, Australia.

99. Department of Pharmacology, National University of Singapore, Singapore.

100. Memory Aging and Cognition Center, National University Health System, Singapore.

101. Institute of Neurogenomics, Helmholtz Zentrum München, German Research Centre for Environmental Health, Neuherberg, Germany.

102. Department of Biomedical Engineering, Illinois Institute of Technology, Chicago, Illinois, USA.

103. Department of Diagnostic Radiology and Nuclear Medicine, Rush University Medical Center, Chicago, Illinois, USA.

104. Academic Unit for Psychiatry of Old Age, University of Melbourne, Victoria, Australia.

105. National Ageing Research Institute, Royal Melbourne Hospital, Melbourne, Victoria, Australia.

106. Harvard Medical School, Boston, Massachusetts, USA.

107. Lurie Center for Autism, Massachusetts General Hospital, Harvard Medical School, Lexington, Massachusetts, USA.

108. Psychiatric and Neurodevelopmental Genetics Unit, Center for Genomic Medicine, Massachusetts General Hospital, Boston, Massachusetts, USA.

109. Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Boston, Massachusetts, USA.

110. Amsterdam Public Health Research Institute, VU Medical Center, Amsterdam, The Netherlands.

111. Department of Psychiatry, University of Oxford, Oxford, UK.

112. NIHR Dementia Biomedical Research Unit, King's College London, London, UK.

113. NORMENT - KG Jebsen Centre for Psychosis Research, Department of Clinical Science, University of Bergen, Bergen, Norway.

114. Dr Einar Martens Research Group for Biological Psychiatry, Center for Medical Genetics and Molecular Medicine, Haukeland University Hospital, Bergen, Norway.

115. Center for integrated Sequencing, iSEQ, Aarhus University, Aarhus, Denmark.

116. Department of Biomedicine, Aarhus University, Aarhus, Denmark.

117. The Lundbeck Foundation Initiative for Integrative Psychiatric Research, iPSYCH, Aarhus and Copenhagen, Denmark.

118. Department of Clinical Neuroscience, Centre for Psychiatric Research, Karolinska Institutet, Stockholm, Sweden.
119. Stockholm Health Care Services, Stockholm County Council, Stockholm, Sweden.
120. UCL Queen Square Institute of Neurology, London, UK
121. Epilepsy Society, Bucks, UK.
122. Centre for Youth Mental Health, The University of Melbourne, Melbourne, Victoria, Australia.
123. Orygen, The National Centre of Excellence in Youth Mental Health, Melbourne, Victoria, Australia.
124. Department of Research and Development, Diakonhjemmet Hospital, Oslo, Norway.
125. Department of Psychology, University of Bath, Bath, United Kingdom.
126. Neuroimaging Unit,Technological Facilities Valdecilla Biomedical Research Institute IDIVAL, Santander, Cantabria, Spain.
127. Avera Institute for Human Genetics, Sioux Falls, South Dakota, USA.
128. Hospital for Sick Children, University of Toronto, Toronto, Ontario, Canada
129. Center for Systems Neuroscience, Boston University, Boston, Massachusetts, USA.
130. Institute of Clinical Chemistry and Laboratory Medicine, University Hospital Regensburg, Regensburg, Germany.
131. Neurodegeneratives Diseases Institute, CNRS UMR 5293, Université de Bordeaux , Bordeaux, France.
132. Pennington Biomedical Research Center, Baton Rouge, Louisiana, USA.
133. Department of Adult Psychiatry, Institute for Clinical Medicine, University of Oslo, Norway.
134. NORMENT - KG Jebsen Centre for Psychosis Research, Oslo University Hospital, Norway.
135. Brain Research Imaging Centre, University of Edinburgh, Edinburgh, UK.

136. Department of Computer Science, Lagos State University, Ojo, Nigeria.

137. Queensland Brain Institute, University of Queensland, Brisbane, Queensland, Australia.

138. Ophthalmology & Visual Sciences Academic Clinical Program (Eye ACP), Duke-NUS Medical School, Singapore.

139. Department of Behavioral Sciences, Rush University Medical Center, Chicago, Illinois, USA.

140. Centre for Healthy Brain Ageing, School of Psychiatry, University of New South Wales, Sydney, New South Wales, Australia.

141. Department of Neurology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

142. Department of Radiology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

143. Department of Psychology, Center for Brain Science, Harvard University, Cambridge, Massachusetts, USA.

144. Section of Gerontology and Geriatrics, Department of Medicine, University of Perugia, Perugia, Italy.

145. Center for Multimodal Imaging and Genetics, University of California, San Diego, California, USA.

146. Department of Cognitive Sciences, University of California, San Diego, California, USA.

147. Department of Neurosciences, University of California, San Diego, California, USA.

148. Department of Psychiatry, University of California, San Diego, California, USA.

149. Department of Radiology, University of California, San Diego, California, USA.

150. Division of Medical Genetics, Department of Biomedicine, University of Basel, Basel, Switzerland.

151. Institute of Human Genetics, University of Bonn, Bonn, Germany.

152. Institute of Neuroscience and Medicine (INM-1), Research Centre Jülich, Jülich, Germany.

153. Reta Lila Weston Institute, UCL Institute of Neurology, London, UK.

154. Department of Molecular Neuroscience, UCL Institute of Neurology, London, UK.

155. Department of ECE, University of New Mexico, Albuquerque, New Mexico, USA.

156. The Mind Research Network & LBERI, Albuquerque, New Mexico, USA.

157. Tri-institutional Center for Translational Research in Neuroimaging and Data Science (TReNDS), Georgia State University, Atlanta, Georgia, USA.

158. Cerebral Imaging Centre, Douglas Mental Health University Institute, Montreal, Québec, Canada.

159. Departments of Psychiatry and Biological and Biomedical Engineering, McGill University, Montreal, QC, Canada.

160. Donders Centre for Cognitive Neuroimaging, Radboud University, Nijmegen, The Netherlands.

161. Institute of Neuroscience and Medicine, Brain & Behaviour (INM-7), Research Centre Jülich, Jülich, Germany.

162. Department of Clinical Neuropsychology, VU University Amsterdam, Amsterdam, The Netherlands.

163. Department of Pediatrics, VU Medical Center, Amsterdam, The Netherlands.

164. Emma Children's Hospital Amsterdam Medical Center, Amsterdam, The Netherlands.

165. Univ. Lille, Labex DISTALZ - U1167 - RID-AGE - Risk factors and molecular determinants of aging-related diseases, Lille, France.

166. Inserm, U1167, Lille, France.

167. Centre Hospitalier Universitaire Lille, Lille, France.

168. Institut Pasteur de Lille, Lille, France.

169. Institute of Diagnostic Radiology and Neuroradiology, University Medicine Greifswald, Greifswald, Germany.

170. Institute for Translational Genomics and Population Sciences, Los Angeles Biomedical Research Institute and Pediatrics at Harbor-UCLA Medical Center, Torrance, California, USA.

171. Institute of Biological Psychiatry, Mental Health Center Sct. Hans, Roskilde, Denmark.

172. Neurogenomics Division, The Translational Genomics Research Institute (TGEN), Phoenix, Arizona, USA.

173. Faculty of Health, Queensland University of Technology (QUT), Brisbane, Queensland, Australia.

174. Institute of Health and Biomedical Innovation, Queensland University of Technology (QUT), Brisbane, Queensland, Australia.

175. Academic Medicine Research Institute, Duke-NUS Medical School, Singapore.

176. Department of Biostatistics, Epidemiology and Informatics, University of Pennsylvania, Philadelphia, Pennsylvania, USA.

177. Dementia Centre for Research Collaboration, UNSW, Sydney, New South Wales, Australia.

178. Department of Neurology, Aristotle University of Thessaloniki, Thessaloniki, Greece.

179. Division of Mind and Brain Research, Department of Psychiatry and Psychotherapy CCM, Charité - Universitätsmedizin Berlin, corporate member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany.

180. Brain Center Rudolf Magnus, Human Neurogenetics Unit, UMC Utrecht, Utrecht, The Netherlands.

181. Department of Molecular and Cellular Therapeutics, the Royal College of Surgeons in Ireland, Dublin, Ireland.

182. Department of Psychiatry, Leiden University Medical Center, Leiden, The Netherlands.

183. Leiden Institute for Brain and Cognition, Leiden University Medical Center, Leiden, The Netherlands.

184. Department of Evolution and Genetics, Dagestan State University, Makhachkala, Dagestan, Russia.

185. Department of Psychiatry, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands.

186. Karakter Child and Adolescent Psychiatry University Center, Nijmegen, The Netherlands.

187. Institute for Community Medicine, Section Epidemiology of Health Care and Community Health, University Medicine Greifswald, Greifswald, Germany.

188. NORMENT - KG Jebsen Centre for Psychosis Research, Department of Psychology, University of Oslo, Oslo, Norway.

189. Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, UK.

190. Translational Neuroscience Laboratory, Department of Psychiatry and Human Behavior, University of California-Irvine, Irvine, California, USA.

191. Center for the Neurobiology of Learning and Memory, University of California Irvine, Irvine, California, USA.

192. University of Wuerzburg, Department of Psychiatry, Psychosomatics and Psychotherapy, Wuerzburg, Germany.

193. Brain and Mind Centre, University of Sydney, Sydney, New South Wales, Australia.

194. Neuroscience Research Australia, Sydney, New South Wales, Australia.

195. University of New South Wales, Sydney, New South Wales, Australia.

196. Department of Internal Medicine, INSERM U 558, University of Toulouse, Toulouse, France.

197. Department of Geriatric Medicine, INSERM U 558, University of Toulouse, Toulouse, France.

198. Department of Psychiatry and Psychotherapy, Charité Universitätsmedizin Berlin, CCM, Berlin, Germany.

199. Center for Neurobehavioral Genetics, University of California, Los Angeles, California, USA.

200. Neurology Division, Beaumont Hospital, Dublin, Ireland.

201. Department of Psychiatry, Carver College of Medicine, University of Iowa, Iowa City, Iowa, USA.

202. Interdepartmental Neuroscience Graduate Program, UCLA School of Medicine, Los Angeles, California, USA.

203. Imaging of Dementia and Aging (IDeA) Laboratory, Department of Neurology, University of California Davis, Davis, California, USA.

204. Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, Massachusetts, USA.

205. NORMENT, KG Jebsen Centre for Psychosis Research, Division of Mental Health and Addiction, Oslo University Hospital & Institute of Clinical Medicine, University of Oslo, Oslo, Norway.

206. School of Mental Health and Neuroscience, Faculty of Health, Medicine and Life Sciences, Maastricht University, Maastricht, The Netherlands.

207. Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Greifswald, Germany.

208. Kaiser Permanent Washington Health Research Institute, Seattle, Washington, USA

209. Department of Epidemiology, University of Washington, Seattle, Washington, USA.

210. Department of Health Services, University of Washington, Seattle, Washington, USA.

211. Institute for Molecular Bioscience, The University of Queensland, Brisbane, Queensland, Australia.

212. Medical and Molecular Genetics, Indiana University School of Medicine, Indianapolis, Indiana, USA.

213. Department of Developmental Disability Neuropsychiatry, School of Psychiatry, UNSW Medicine, Sydney, New South Wales, Australia.

214. Biomedical Research Unit for Dementia, King's College London, London, UK.

215. Department of Neuroimaging, Institute of Psychiatry, King's College London, London, UK.

216. Division of Clinical Geriatrics, Department of Neurobiology, Care Sciences and Society, Karolinska Institute, Stockholm, Sweden.

217. Division of Genetics, Department of Medicine, Brigham and Women's Hospital, Boston, Massachusetts, USA.

218. Section of Computational Biomedicine, Department of Medicine, Boston University School of Medicine, Boston, Massachusetts, USA.

219. Department of Radiology, Mayo Clinic, Rochester, Minnesota, USA.

220. School of Medical Sciences, UNSW, Sydney, New South Wales, Australia.

221. Department of Biomedicine, University of Basel, Basel, Switzerland.

222. C. & O. Vogt Institute for Brain Research, Heinrich Heine University Düsseldorf, Düsseldorf, Germany.

223. Department of Psychiatry and Human Behavior, University of California-Irvine, Irvine, California, USA.

224. Laboratory of Neuro Imaging, USC Mark and Mary Stevens Neuroimaging and Informatics Institute, Keck School of Medicine of the University of Southern California, Los Angeles, California, USA.

225. Umeå Centre for Functional Brain Imaging (UFBI), Umeå University, Umeå 901 87, Sweden.

226. Radiation Sciences, Umeå University, Umeå, Sweden.

227. Department of Radiology, Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts, USA.

228. Department of Internal Medicine, Erasmus Medical Center, Rotterdam, The Netherlands.

229. Department of Neuroscience, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

230. Institute of Genetic Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

231. Department of Psychiatry, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

232. Imaging Physics, Faculty of Applied Sciences, Delft University of Technology, The Netherlands.

233. Institut Pasteur, Paris, France.

234. Department of Neurology, University of Washington, Seattle, Washington, USA.

235. Department of Neurology, Erasmus MC, Rotterdam, The Netherlands.

236. Cognitive Neuroscience Center, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands.

237. Bloorview Research Institute, Holland Bloorview Kids Rehabilitation Hospital, Toronto, Ontario, Canada.

238. Department of Psychology, University of Toronto, Toronto, Ontario, Canada.

239. Department of Psychiatry, University of Toronto, Toronto, Ontario, Canada.

240. Clinic for Cognitive Neurology, University Clinic Leipzig, Leipzig, Germany.

241. HMNC Brain Health, Munich, Germany.

242. Department of Neurology, Hopital Erasme, Universite Libre de Bruxelles, Brussels, Belgium.

243. Department of Psychology, Georgia State University, Atlanta, Georgia, USA.

244. Department of Medical Genetics, Oslo University Hospital, Oslo, Norway.

245. Munich Cluster for Systems Neurology (SyNergy), Munich, Germany.

246. Institute of Translational Medicine, University of Liverpool, Liverpool, UK.

247. Institute of Clinical Medicine, Neurology, University of Eastern Finland, Kuopio, Finland.

248. Neurocentre Neurology, Kuopio University Hospital, Kuopio, Finland.

249. INSERM, Research Unit 1000 “Neuroimaging and Psychiatry”, Paris Saclay University, Paris Descartes University; DIGITEO Labs, Gif sur Yvette, France.

250. Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald, Greifswald, Germany.

251. German Center for Cardiovascular Research (DZHK eV), partner site Greifswald, Greifswald, Germany.

252. Department of Neuroinformatics, Araya, Inc., Tokyo, Japan.

253. Institute of Cognitive Neuroscience, University College London, London, UK.

254. School of Psychology, University of Sussex, Brighton, UK.

255. Department of Neurobiology, Care Sciences and Society, Karolinska Institutet, Stockholm, Sweden.

256. Department of Child and Adolescent Psychiatry/Psychology, Erasmus MC-Sophia Children's Hospital, Rotterdam, The Netherlands.

257. Department of Clinical Genetics, Maastricht University Medical Center, Maastricht, The Netherlands.

258. GROW School for Oncology and Developmental Biology, Maastricht, The Netherlands.

259. Centre for Advanced Imaging, University of Queensland, Brisbane, Queensland, Australia.

260. Department of Genomics, Life & Brain Center, University of Bonn, Germany.

261. Neuropsychiatric Institute, Prince of Wales Hospital, Sydney, New South Wales, Australia.

262. Department of Psychology, VU University Amsterdam, Amsterdam, The Netherlands.

263. Center for Imaging of Neurodegenerative Disease, San Francisco VA Medical Center, University of California, San Francisco, USA.

264. Department of Radiology and Biomedical Imaging, University of California, San Francisco, California, USA.

265. Leiden Academic Centre for Drug Research (LACDR), Leiden University, The Netherlands.

266. Human Genetics Center, University of Texas Health Science Center at Houston, Houston, Texas, USA.

267. UK Dementia Research Institute, University of Edinburgh, Edinburgh, UK.

268. Department of Neurology, Center for Neuroscience, University of California at Davis, Sacramento, California, USA.

269. Medical and Population Genetics, Broad Institute, Cambridge, Massachusetts, USA.

270. Center for Translational & Computational Neuroimmunology, Department of Neurology, Columbia University Medical Center, New York, New York, USA.

271. Department of Neuroscience, Baylor College of Medicine, Houston, Texas, USA.

272. Department of Molecular & Human Genetics, Baylor College of Medicine, Houston, Texas, USA.

273. Jan and Dan Duncan Neurological Research Institute, Texas Children's Hospital, Houston, Texas, USA.

* CLS, HHHA, DPH, and CCW contributed equally.

** SEM, JMS, PMT, SS, and MAI jointly supervised this work.

Corresponding Authors:

Dr. Claudia L. Satizabal

Glenn Biggs Institute for Alzheimer's & Neurodegenerative Diseases

7703 Floyd Curl Drive, MSC 8070, San Antonio, TX 78229

Phone: +1 210-450-8417

Email: satizabal@uthscsa.edu

Dr. M. Arfan Ikram

Erasmus University Medical Center Rotterdam

P.O. Box 2040, 3000 CA, Rotterdam, The Netherlands

Phone: +31 10 7043930

E-mail: m.a.ikram@erasmusmc.nl

Abstract

Subcortical brain structures are integral to motion, consciousness, emotions, and learning. We identified common genetic variation related to the volumes of nucleus accumbens, amygdala, brainstem, caudate nucleus, globus pallidus, putamen, and thalamus, using genome-wide association analyses in almost 40,000 individuals from CHARGE, ENIGMA and the UK-Biobank. We show that variability in subcortical volumes is heritable, and identify 48 significantly associated loci (40 novel at the time of analysis). Annotation of these loci utilizing gene expression, methylation, and neuropathological data identified 199 genes putatively implicated in neurodevelopment, synaptic signaling, axonal transport, apoptosis, inflammation/infection, and susceptibility to neurological disorders. This set of genes is significantly enriched for *Drosophila* orthologs associated with neurodevelopmental phenotypes, suggesting evolutionarily conserved mechanisms. Our findings uncover novel biology and potential drug targets underlying brain development and disease.

Subcortical brain structures are essential for the control of autonomic and sensorimotor functions[1,2], modulation of processes involved in learning, memory, and decision-making[3,4], as well as in emotional reactivity[5,6] and consciousness[7]. They often act through networks influencing input to and output from the cerebral cortex[8,9]. The pathology of many cognitive, psychiatric, and movement disorders is restricted to, begins in, or predominantly involves subcortical brain structures and related circuitries[10]. For instance, tau pathology has shown to manifest itself early in the brainstem of individuals with Alzheimer's disease before spreading to cortical areas through efferent networks[11]. Similarly, the formation of Lewy bodies and Lewy neurites in Parkinson's disease appears early in the lower brainstem (and olfactory structures) before affecting the substantia nigra[12].

Recent investigations have identified genetic loci influencing the volumes of the putamen, caudate, and pallidum, which pointed to genes controlling neurodevelopment and learning, apoptosis, and transport of metals[13,14]. However, a larger study combining these samples, which include individuals of a broad age-range across diverse studies, would enable increased power to identify additional novel genetic variants contributing to variability in subcortical structures, and further improve our understanding of brain development and disease.

We sought to identify novel genetic variants influencing the volumes of seven subcortical structures (nucleus accumbens, amygdala, caudate nucleus, putamen, globus pallidus, thalamus, and brainstem – including mesencephalon, pons, and medulla oblongata), through genome-wide association (GWA) analyses in almost 40,000 individuals from 53

study samples (Supplementary Table 1-3) from the Cohorts of Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium, the Enhancing Neuro Imaging Genetics through Meta-Analysis (ENIGMA) consortium, and the United Kingdom Biobank (UKBB).

RESULTS

Heritability

To examine the extent to which genetic variation accounts for variation in subcortical brain volumes, we estimated their heritability in two family-based cohorts: the Framingham Heart Study (FHS) and the Austrian Stroke Prevention Study (ASPS-Fam). Our analyses are in line with previous studies conducted in twins[15], suggesting that variability in subcortical volumes is moderately to highly heritable. The structures with highest heritability in the FHS and the ASPS-Fam are the brainstem (ranging from 79-86%), caudate nucleus (71-85%), putamen (71-79%) and nucleus accumbens (66%); followed by the globus pallidus (55-60%), thalamus (47-54%), and amygdala (34-59%) (Figure 1, Supplementary Table 4). We additionally estimated SNP-based heritability using GCTA in the Rotterdam Study, and LD score regression (LDSC) in the full European sample. As expected, SNP-based heritability estimates were somewhat lower, ranging from 47% for the thalamus to 17% for the amygdala using GCTA, and ranging from 33% for the brainstem to 9% for the amygdala using LDSC. These values are consistent with heritability estimates reported by the UKBB[14].

Genome-wide associations

We undertook a GWA analysis on the MRI-derived volumes of subcortical structures using the 1000 Genomes Project[16] reference panel (phase 1 v.3) for imputation of missing variants in CHARGE and ENIGMA. The UKBB performed imputation of variants using the HRC reference panel[17] (see details on image acquisition and genotyping in Supplementary Table 5 and Supplementary Table 6, respectively). Our sample comprised up to n = 37,741 individuals of European ancestry from 48 study samples across CHARGE, ENIGMA and the UKBB. Additionally, we included three samples for generalization in African-Americans (up to n = 769), and two for generalization in Asians (n = 341). Details on the population characteristics, definition of the outcome and genotyping can be found in the supplement (Supplementary Tables 2-5). Each study examined the association of genetic variants with minor allele frequency (MAF) ≥1% to the volumes of subcortical structures (average volume for bilateral structures) using additive genetic models adjusted for sex, age, total intracranial volume (or total brain volume in the UKBB); as well as age^2, population structure, psychiatric diagnosis (ENIGMA cohorts), and study site when applicable. After quality control, we conducted meta-analyses per ethnicity combining all samples using sample-size-weighted fixed effects methods in METAL[18]. An analysis of genetic correlations showed consistency of associations across the CHARGE-ENIGMA and the UKBB ($r_g > 0.94$; $P < 1.46 \times 10^{-15}$), demonstrating the similar genetic architecture of subcortical volumes in these two datasets.

We identified 48 independent genome-wide significant single nucleotide polymorphisms (SNPs) across all seven subcortical structures, 40 of which are novel at the

time of analysis (Table 1). Among these, 26 SNPs were located within genes (one missense, 25 intronic), and 22 in intergenic regions. Most of the inflation observed in the quantile plots (Supplementary Figure 1) is due to polygenic effects. We carried forward these 48 SNPs for *in-silico* generalization in African-American and Asian samples, and performed a combined meta-analysis of all samples (Supplementary Table 7). Of the 46 SNPs present in the generalization samples, the direction of association was the same for 13 across all ethnicities and for an additional 6 SNPs in either the African-American or the Asian samples. In the combined meta-analysis, 43 of the 48 associations remained significant, and for 21 SNPs, the strength of association increased when all samples were combined. Although we did not find significant associations for most SNPs at the generalization sample level, likely due to their limited sample size, the sign test for the direction of effect suggested that a large proportion of the SNPs associated with subcortical volumes in the European sample are also associated in the African-American and Asian samples at the polygenic level ($P < 1 \times 10^{-4}$; Supplementary Table 8).

To functionally annotate the 48 SNPs identified in the European sample, we used Locus Zoom[19], investigated expression quantitative trait loci (eQTL) and methylation QTL (meQTL) in post-mortem brains from the Religious Order Study and the Rush Memory and Aging Project (ROSMAP), and also queried *cis*- and *trans*-eQTL datasets in brain and non-brain tissues for the top 48 SNPs or their proxies ($r^2>0.8$), using the European population reference (Supplementary Tables 9-12). Lead variants and their proxies were annotated to genes based on the combination of physical proximity, eQTL and meQTL, which in some instances assigned more than one gene to a single SNP. Most of our index SNPs had genes

assigned based on more than one functional source. This strategy allowed us to identify 199 putatively associated genes (Supplementary Table 13). More details can be found in the Supplementary note.

Associations with cognition and neuropathology

Although individual SNPs were not related to neuro-pathological traits or cognitive function in ROSMAP (Supplementary Table 14), we found that cortical mRNA expression of 12 of our putatively associated genes was associated with neuropathological alterations typically observed in Alzheimer's Disease (Supplementary Table 15). These included β-amyloid load / presence of neuritic plaques (*APOBR, FAM65C, KTN1, NUPR1, OPA1*) and tau density / neurofibrillary tangles (*FAM65C, MEPCE, OPA1, STAT1*). Many of these genes, together with *ANKRD42, BCL2L1, RAET1G, SGTB,* and *ZCCHC14*, were also related to cognitive function.

Phenotypic and genetic correlations

We explored both phenotypic (Supplementary Table 16) and genetic (Supplementary Table 17) correlations among subcortical volumes. We also investigated genetic correlations of subcortical volumes with traits previously examined in the CHARGE and ENIGMA consortia, including MRI-defined brain volumes[20,21,22], stroke subtypes[23], anthropometric traits[24], general cognitive function[25], Alzheimer's disease[26], Parkinson's Disease[27], bipolar disorder and schizophrenia[28], and attention deficit/hyperactivity disorder (ADHD)[29]. We observed strong phenotypic and genetic overlap among most

126 subcortical structures using LDSC methods, consistent with our finding that many of the
127 loci identified have pleiotropic effects on the volumes of several subcortical structures.
128 As expected, we found strong genetic correlations among the nuclei composing the
129 striatum, particularly for nucleus accumbens with caudate nucleus ($P = 9.83 \times 10^{-19}$), and
130 with putamen ($P = 1.02 \times 10^{-17}$). The genetic architecture of thalamic volume highly
131 overlapped with that of most subcortical volumes, except for the caudate nucleus. In
132 contrast, there were no significant genetic correlations for the volume of the brainstem
133 with that of most structures, with the exception of very strong correlations with volumes of
134 the thalamus ($P = 1.56 \times 10^{-22}$) and the globus pallidus ($P = 1.52 \times 10^{-21}$). Individual level
135 analyses using GCTA in the Rotterdam Study (n = 3,486) showed similar correlations
136 despite the smaller sample.
137 We also observed strong genetic correlations for hippocampal volumes with amygdalar
138 and thalamic volumes. Height correlated with thalamic volumes and volume of the
139 brainstem was inversely correlated with ADHD. Notably, caudate nucleus volumes
140 correlated with white matter hyperintensity burden.
141
142 ***Cross-species analysis***
143 To investigate for potential evolutionarily conserved requirements of our gene-set in
144 neurodevelopment, neuronal maintenance, or both, we examined available genetic and
145 phenotypic data from the fruit fly, *Drosophila melanogaster*. Importantly, compared to
146 mammalian models, the fly genome has been more comprehensively interrogated for roles
147 in the nervous system. We found that a large proportion of candidate genes for human

148 subcortical volumes are strongly conserved in the *Drosophila* genome (59%), and many of
149 these genes appear to have conserved nervous system requirements (Supplementary Table
150 18). To examine if this degree of conservation was greater than that expected by chance,
151 we leveraged systematic, standardized phenotype data based on FlyBase annotations using
152 controlled vocabulary terms. Indeed, 22% of the conserved fly homologs are documented
153 to cause "neuroanatomy defective" phenotypes in flies, representing a significant ($P = 7.3 \times$
154 10^{-4}), nearly two-fold enrichment compared to 12.9% representing all *Drosophila* genes
155 associated with such phenotypes (Supplementary Table 19).
156
157 ***Partitioning heritability***
158 We further investigated enrichment for functional categories of the genome using
159 stratified LDSC methods[30] (Figure 2). Super enhancers were significantly enriched in most
160 subcortical structures, with 17% of SNPs explaining 43% of SNP-heritability in the
161 brainstem, 39% in the caudate, 44% in the pallidum, 37% in the putamen, and 38% in the
162 thalamus. Similarly, strong enrichment was observed for regular enhancers (H3K27ac
163 annotations from Hnisz[31]) in several subcortical structures, explaining over 60% of their
164 SNP-heritability. Conserved regions were enriched in the nucleus accumbens and the
165 brainstem, with 2.6% of SNPs explaining 53% and 35% of their SNP heritability,
166 respectively. Finally, only the brainstem showed enrichment for transcription start sites
167 (TSS), with 1.8% of SNPs explaining 26% of this structure SNP-heritability. Full results are
168 presented in Supplementary Table 20.
169

Protein-protein interactions

To explore potential functional relationships between proteins encoded by our set of genes, we conducted protein-protein interaction analyses in STRING[32]. Our results showed enrichment of genes involved in brain-specific pathways (i.e. regulation of neuronal death and neuronal apoptosis), as well as immune-related (i.e. antigen processing, Epstein-Barr virus infection) and housekeeping processes (i.e. proteasome, cell differentiation, signaling). Figure 3 shows these protein networks, and the detailed pathways are presented in Supplementary Table 21.

DISCUSSION

We undertook the largest GWA meta-analysis of variants associated with MRI-derived volumes of the nucleus accumbens, amygdala, brainstem, caudate nucleus, globus pallidus, putamen, and thalamus; in almost 40,000 individuals from 53 study samples worldwide. Our analyses identified a set of 199 candidate genes influencing the volume of these subcortical brain structures, most of which have relevant roles in the nervous system.

Our results show wide overlap of genetic variants determining the volume of subcortical structures as elucidated from genetic correlations and individual look-ups among structures. We find that 26 candidate genes may influence more than one structure. For instance, significant SNPs near *KTN1,* are also associated with the volume of the nucleus accumbens, caudate nucleus, and globus pallidus, suggesting that this genomic region may have an important role in determining multiple subcortical brain volumes during development. Furthermore, 14 of the candidate genes were associated with the caudate, globus pallidus and putamen, supporting the shared genetic architecture of the functionally defined corpus striatum.

We identified genes implicated in ***neurodevelopment***. We confirm the 11q14.3 genomic region near the *FAT3* gene, previously associated with the caudate nucleus[13], additionally associated with the putamen in our analysis. This gene encodes a conserved cellular adhesion molecule implicated in neuronal morphogenesis and cell migration based on mouse genetic studies[33]. SNPs near *PBX3* were associated with caudate volume. *PBX3* is robustly expressed in the developing caudate nucleus of the non-human primate, *Macaca fuscata*, consistent with a role in striatal neurogenesis[34].

We found several genes involved in insulin/IGF1 signaling, including *IGF1*, *PAPPA, GRB10, SH2B1* and *TXNDC5* across the amygdala, brainstem, caudate, and putamen. *PAPPA* encodes a secreted metalloproteinase that cleaves IGFBPs, thereby releasing bound IGF. Although IGF may be beneficial in early- and midlife, its effects may be detrimental during aging. Studies of PAPPA similarly support antagonistic pleiotropy. Low circulating PAPPA levels are a marker for adverse outcomes in human embryonic development[35], but in later life, higher levels have been associated with acute coronary syndromes and heart failure[36,37]. Further, Grb10 and SH2B1 act as regulators of insulin/IGF1 signaling through their SH2 domains[38]. Finally, *TXNDC5* has been suggested to increase IGF1 activity by inhibiting the expression IGFBP1 in the context of rheumatoid arthritis[39].

Additional genes related to neurodevelopment include *PTPN1* (brainstem), *ALPL* and *NBPF3,* (both related to the globus pallidus), and *SLC20A2* (nucleus accumbens). In studies of both human and mouse embryonic stem cells, *PTPN1* was implicated as a critical regulator of neural differentiation[40]. In addition, *PTPN1* encodes a target for the transcriptional regulator encoded by *MECP2*, which causes the neurodevelopmental disorder Rett Syndrome, and inhibition of *PTPB1* is being explored as a therapeutic strategy in mouse Rett models[41]. *ALPL* mediates neuronal differentiation early during development and post-natal synaptogenesis in transgenic mouse models[42]. ALPL may also help propagate the neurotoxicity induced by tau[43], and its activity increases in Alzheimer's disease[44] and cognitive impairment[45]. *NBPF3* belongs to the neuroblastoma breakpoint family, which encodes domains of the autism- and schizophrenia-related DUF1220 protein[46]. *SLC20A2*, related to the globus pallidus and the thalamus, encodes an inorganic

phosphate transporter for which more than 40 mutations have been described in association with familial idiopathic basal ganglia calcification (Fahr's Syndrome)[47,48]. It is interesting to note that other three solute carrier genes were identified in this GWA (*SLC12A9, SLC25A29, SLC39A8*), suggesting that the molecular transport of metals, amino acids, and other solutes across the cellular membrane could play an important role in the development of subcortical brain structures.

Several genes were related to ***synaptic signaling pathways***. We found a SNP in *NP*TX1 related to the thalamus, a gene expressed in the nervous system which restricts synapse plasticity[49], and induces β-amyloid neurodegeneration in human and mouse brain tissues[50]. Additionally, the identified an intronic SNP in *SGTB* for the brainstem, which was an eQTL for the expression of SGTB in dorsolateral prefrontal cortex. Experimental rat models showed that βSGT, highly expressed in brain, forms a complex with the cysteine string protein and heat-shock protein cognate (CSP/Hsc70) complex to function as a chaperone guiding the refolding of misfolded proteins near synaptic vesicles[51]. Other experimental studies in *C. elegans*, showed that the genetic manipulation of the ortholog, *sgt-1*, suppresses toxicity associated with expression of the human β-amyloid peptide[52]. Other genes involved in synaptic signaling are *CHPT1* (brainstem), involved in phosphatidylcholine metabolism in the brain; *KATNA1*(brainstem), a conserved regulator of neuronal process formation, outgrowth, and synaptogenesis[53,54]; and *DLG2* (putamen), encoding an evolutionarily conserved scaffolding protein involved in glutamatergic-mediated synaptic signaling and cell polarity[55] that has been associated with schizophrenia[56], cognitive impairment[57], and Parkinson's disease[58].

Another set of SNPs point to genes involved in ***autophagy and apoptotic processes***, such as *DRAM1* and *FOXO3*, both related to brainstem volumes. *DRAM1* encodes a lysosomal membrane protein involved in activating TP53-mediated autophagy and apoptosis,[59] and mouse models mimicking cerebral ischemia and reperfusion have found that inhibiting the expression of *DRAM1* worsens cell injury[60]. The top SNP was also associated with a CpG site proximate to active TSS upstream of *DRAM1* in several mature brain tissues (S3.6). *FOXO3* has been recently identified as pivotal in an astrocyte network conserved across humans and mice involved in stress, sleep, and Huntington's disease[61], and has been related to longevity[62]. In *Drosophila*, a *FOXO3* ortholog regulates dendrite number and length in the peripheral nervous system[63], and in the zebrafish, *Danio rario*, *Foxo3a* knockdown led to apoptosis and mispatterning of the embryonic CNS[64]. Additional genes involved in apoptotic processes are *BCL2L1* (globus pallidus and putamen), *BIRC6* (globus pallidus) and *OPA1* (brainstem).

Other genes have been implicated in ***axonal transport***. We confirm the association between the 13q22 locus near *KTN1* with putamen volume[13]and expand by showing that this region is also associated with the nucleus accumbens, caudate and the globus pallidus . The most significant SNP (rs945270) is a robust eQTL for *KTN1* in peripheral blood cells. This gene encodes a kinesin-binding protein involved in the transport of cellular components along microtubules[65], and impairment of these molecular motors has been increasingly recognized in neurological diseases with a subcortical component[66]. The 5q12 locus upstream from *MAST4* was associated with nucleus accumbens volume. *MAST4* encodes a member of the microtubule-associated serine/threonine kinases. This gene has

been associated with hippocampal volumes[20] and juvenile myoclonic epilepsy[67],and it appears to be differentially expressed in the prefrontal cortex of atypical cases of frontotemporal lobar degeneration[68]. In *Drosophila*, the knockdown of a conserved *MAST4* homolog enhanced the neurotoxicity of human tau[69], which aggregates to form neurofibrillary tangle pathology in Alzheimer's disease. Further, we identified SNPs near *NEFL* and *NEFM* (globus pallidus), where the top SNP was an eQTL for these genes in subcortical brain tissue and esophagus mucosa. *NEFL* encodes the light chain, and NEFM the medium chain of the neurofilament. These proteins determine neuronal caliber and conduction velocity[70]. Mutations in NEFL/M genes have been related to neuropsychiatric disorders and both proteins are increasingly recognized as powerful biomarkers of neurodegeneration[71].

Finally, several of our candidate genes are also involved in ***inflammation, immunity and infection*** (*ANKRD42, DEFB124, IL27, NLRC4, PILRA/B, TRIM23, TRIM4*), in line with the PPI analysis highlighting the KEGG-Epstein-Barr virus infection pathway. This suggests that immune-related processes may be an important determinant influencing subcortical volumes, as has been shown by other GWAS of neurologic traits[72,73].

Overall, the loci identified by our study pinpoint candidate genes not only associated with human subcortical brain volumes, but also reported to disrupt invertebrate neuroanatomy when manipulated in *Drosophila* and many other animal models. Thus, our results are in line with the knowledge that the genomic architecture of central nervous system development has been strongly conserved during evolution. Partitioning

heritability results suggest the nucleus accumbens and the brainstem are particularly enriched in conserved regions.

One of the main limitations of our study was the small size of our generalization samples, which limits the generalizability of our results to non-European ethnicities. However, our analyses suggest significant concordance for the direction of effect across all ethnicities at the polygenic level. We hope diverse samples become increasingly available to further confirm our findings and make new discoveries. Additionally, we have focused on the discovery of common and less frequent variants. Further efforts to also reveal rare variants and epigenetic signatures associated with subcortical structures will provide an even more refined understanding of the underlying mechanisms involved.

In conclusion, we describe multiple genes associated with the volumes of MRI-derived subcortical structures in a large sample, leveraging diverse bioinformatic resources to validation and follow-up our findings. Our analyses indicate that the variability of evolutionarily old subcortical volumes of humans is moderately to strongly heritable, and that their genetic variation is also strongly conserved across different species. The majority of the variants identified in this analysis point to genes involved in neurodevelopment, regulation of neuronal apoptotic processes, synaptic signaling, axonal transport, inflammation/immunity, and susceptibility to neurological disorders. We show that the genetic architecture of subcortical volumes overlaps with that of anthropometric measures and neuropsychiatric disorders. In summary, our findings greatly expand current

understanding of the genetic variation related to subcortical structures, which can help identify novel biological pathways of relevance to human brain development and disease.

ACKNOWLEDGEMENTS

We thank all study participants for their contributions to make this research possible. Full acknowledgements and grant support details are provided in the Supplementary Note.

AUTHOR CONTRIBUTIONS

CLS drafted the manuscript with contributions from HHHA, DPH, CCW, TVL, AAV, SE, AKH, MWV, DJ, TGMVE, CDW, MJW, SEF, KAM, PJH, BF, HJG, ADJ, OLL, SDe, SEM, JMS, PMT, SS, and MAI.

MS, NJ, LRY, TVL, GC, LA, MER, ADB, IK, MA, SA, SE, RRS, AKH, HJJ, AS, JB, MWV, AVW, KW, NA, SH, ALG, PHL, SG, SLH, DK, LS, SML, IA, EW, DTG, JCI, LNV, RB, FC, DJ, OC, UKH, BSA, CYC, AAA, MPB, AFM, SKM, PA, AJS, DCML, TYW, LSh, PGS, EJCdG, MT, KRVE, NJAVd, AMM, JSR, NR, WH, MCVH, JBJK, LMOL, AHo, GH, MBa, SR, JJHo, ASi, NH, PRS, TWM, PMa, OGru, NAG, JES, HLe, BM, DVR, IJD, RMB, IM, RK, HV, MJW, DvtE, MMN, SEF, ASB, KAM, NRS, DJH, HJG, CMvD, JMW, CDe, PLDJ, and VG contributed to the preparation of data; CLS, HHHA, DPH, MJK, JLS, MS, MSa, NJ, GVR, AVS, JCB, XJ, ML, EH, AT, SJvdL, JY, LRY, SL, KJY, GC, MER, NJA, HJJ, AVW, SH, NMS, SG, DTG, JS, CYC, LMOL, QY, ATh, IOF, DvtE, CDe, and PLDJ performed statistical analyses; and CLS, HHHA, CCW, MJK, TVL, SL, YH, KJY, JDE, QY, and ADJ carried out downstream analyses.

SEM, JMS, PMT, SS, and MAI jointly supervised this work.

All authors reviewed the manuscript for intellectual content.

COMPETING INTERESTS

DPH is currently an employee at Genentech, Inc. DJ has received travel and speaker's honoraria from Janssen-Cilag and research funding from DFG. RLB is a consultant for Pfizer, Roche. PA is a scientific adviser for Genoscreen. TYW is a consultant & advisory board member for Allergan, Bayer, Boehringer-Ingelheim, Genentech, Merck, Novartis, Oxurion (formerly ThromboGenics), Roche; and is a co-founder of Plano and EyRiS. AMM has received grant support from Eli Lilly, Janssen, Pfizer, and the Sackler Trust. BMP serves on the Steering Committee of the Yale Open Data Access Project funded by Johnson & Johnson. AML is a member of the advisory board for the Lundbeck Int. Neuroscience Foundation and Brainsway; is a member of the editorial board for the American Association for the Advancement of Science and Elsevier; is a faculty member of the Lundbeck International Neuroscience Foundation; and is a consultant for Boehringer Ingelheim. WJN is founder, scientific lead and shareholder of Quantib BV. MMN is a shareholder of the Life & Brain GmbH and receives a salary from Life & Brain GmbH; has received support from Shire for attending conferences; and has received financial remuneration from the Lundbeck Foundation, the Robert Bosch Foundation and the Deutsches Ärzteblatt for participation in scientific advisory boards. BF has received educational speaking fees from Shire and Medice. HJG has received travel grants and speaker's honoraria from Fresenius Medical Care, Neuraxpharm and Janssen Cilag, as well as research funding from Fresenius Medical Care.

REFERENCES

1. Marsden, C.D. The mysterious motor function of the basal ganglia: the Robert Wartenberg Lecture. *Neurology* **32**, 514-39 (1982).
2. Yin, H.H. & Knowlton, B.J. The role of the basal ganglia in habit formation. *Nat Rev Neurosci* **7**, 464-76 (2006).
3. McDonald, A.J. & Mott, D.D. Functional neuroanatomy of amygdalohippocampal interconnections and their role in learning and memory. *J Neurosci Res* (2016).
4. Hikosaka, O., Kim, H.F., Yasuda, M. & Yamamoto, S. Basal ganglia circuits for reward value-guided behavior. *Annu Rev Neurosci* **37**, 289-306 (2014).
5. Salzman, C.D. & Fusi, S. Emotion, cognition, and mental state representation in amygdala and prefrontal cortex. *Annu Rev Neurosci* **33**, 173-202 (2010).
6. Floresco, S.B. The nucleus accumbens: an interface between cognition, emotion, and action. *Annu Rev Psychol* **66**, 25-52 (2015).
7. Fabbro, F., Aglioti, S.M., Bergamasco, M., Clarici, A. & Panksepp, J. Evolutionary aspects of self- and world consciousness in vertebrates. *Front Hum Neurosci* **9**, 157 (2015).
8. Alexander, G.E., DeLong, M.R. & Strick, P.L. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu Rev Neurosci* **9**, 357-81 (1986).
9. Jahanshahi, M., Obeso, I., Rothwell, J.C. & Obeso, J.A. A fronto-striato-subthalamic-pallidal network for goal-directed and habitual inhibition. *Nat Rev Neurosci* **16**, 719-32 (2015).
10. Shepherd, G.M. Corticostriatal connectivity and its role in disease. *Nat Rev Neurosci* **14**, 278-91 (2013).
11. Stratmann, K. *et al.* Precortical Phase of Alzheimer's Disease (AD)-Related Tau Cytoskeletal Pathology. *Brain Pathol* **26**, 371-86 (2016).

12. Del Tredici, K., Rub, U., De Vos, R.A., Bohl, J.R. & Braak, H. Where does Parkinson disease pathology begin in the brain? *J Neuropathol Exp Neurol* **61**, 413-26 (2002).
13. Hibar, D.P. *et al.* Common genetic variants influence human subcortical brain structures. *Nature* **520**, 224-9 (2015).
14. Elliott, L.T. *et al.* Genome-wide association studies of brain imaging phenotypes in UK Biobank. *Nature* **562**, 210-216 (2018).
15. Renteria, M.E. *et al.* Genetic architecture of subcortical brain regions: common and region-specific genetic contributions. *Genes Brain Behav* **13**, 821-30 (2014).
16. Clarke, L. *et al.* The 1000 Genomes Project: data management and community access. *Nat Methods* **9**, 459-62 (2012).
17. McCarthy, S. *et al.* A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet* **48**, 1279-83 (2016).
18. Willer, C.J., Li, Y. & Abecasis, G.R. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* **26**, 2190-1 (2010).
19. Pruim, R.J. *et al.* LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics* **26**, 2336-7 (2010).
20. Hibar, D.P. *et al.* Novel genetic loci associated with hippocampal volume. *Nat Commun* **8**, 13624 (2017).
21. Adams, H.H. *et al.* Novel genetic loci underlying human intracranial volume identified through genome-wide association. *Nat Neurosci* **19**, 1569-1582 (2016).
22. Verhaaren, B.F. *et al.* Multiethnic genome-wide association study of cerebral white matter hyperintensities on MRI. *Circ Cardiovasc Genet* **8**, 398-409 (2015).
23. Malik, R. *et al.* Multiancestry genome-wide association study of 520,000 subjects identifies 32 loci associated with stroke and stroke subtypes. *Nat Genet* **50**, 524-537 (2018).

24. Yengo, L. *et al.* Meta-analysis of genome-wide association studies for height and body mass index in approximately 700000 individuals of European ancestry. *Hum Mol Genet* **27**, 3641-3649 (2018).
25. Davies, G. *et al.* Study of 300,486 individuals identifies 148 independent genetic loci influencing general cognitive function. *Nat Commun* **9**, 2098 (2018).
26. Kunkle, B.W. *et al.* Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates Abeta, tau, immunity and lipid processing. *Nat Genet* **51**, 414-430 (2019).
27. Simon-Sanchez, J. *et al.* Genome-wide association study reveals genetic risk underlying Parkinson's disease. *Nat Genet* **41**, 1308-12 (2009).
28. Bipolar, D., Schizophrenia Working Group of the Psychiatric Genomics Consortium. Electronic address, d.r.v.e., Bipolar, D. & Schizophrenia Working Group of the Psychiatric Genomics, C. Genomic Dissection of Bipolar Disorder and Schizophrenia, Including 28 Subphenotypes. *Cell* **173**, 1705-1715 e16 (2018).
29. Demontis, D. *et al.* Discovery of the first genome-wide significant risk loci for attention deficit/hyperactivity disorder. *Nat Genet* **51**, 63-75 (2019).
30. Finucane, H.K. *et al.* Partitioning heritability by functional annotation using genome-wide association summary statistics. *Nat Genet* **47**, 1228-35 (2015).
31. Hnisz, D. *et al.* Super-enhancers in the control of cell identity and disease. *Cell* **155**, 934-47 (2013).
32. Szklarczyk, D. *et al.* STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res* **43**, D447-52 (2015).
33. Deans, M.R. *et al.* Control of neuronal morphology by the atypical cadherin Fat3. *Neuron* **71**, 820-32 (2011).

34. Takahashi, K. *et al.* Expression of FOXP2 in the developing monkey forebrain: comparison with the expression of the genes FOXP1, PBX3, and MEIS2. *J Comp Neurol* **509**, 180-9 (2008).

35. Kjaer-Sorensen, K. *et al.* Pregnancy-associated plasma protein A (PAPP-A) modulates the early developmental rate in zebrafish independently of its proteolytic activity. *J Biol Chem* **288**, 9982-92 (2013).

36. Bayes-Genis, A. *et al.* Pregnancy-associated plasma protein A as a marker of acute coronary syndromes. *N Engl J Med* **345**, 1022-9 (2001).

37. Funayama, A. *et al.* Serum pregnancy-associated plasma protein a in patients with heart failure. *J Card Fail* **17**, 819-26 (2011).

38. Desbuquois, B., Carre, N. & Burnol, A.F. Regulation of insulin and type 1 insulin-like growth factor signaling and action by the Grb10/14 and SH2B1/B2 adaptor proteins. *FEBS J* **280**, 794-816 (2013).

39. Li, J. *et al.* TXNDC5 contributes to rheumatoid arthritis by down-regulating IGFBP1 expression. *Clin Exp Immunol* **192**, 82-94 (2018).

40. Matulka, K. *et al.* PTP1B is an effector of activin signaling and regulates neural specification of embryonic stem cells. *Cell Stem Cell* **13**, 706-19 (2013).

41. Krishnan, N. *et al.* PTP1B inhibition suggests a therapeutic strategy for Rett syndrome. *J Clin Invest* **125**, 3163-77 (2015).

42. Sebastian-Serrano, A. *et al.* Tissue-nonspecific Alkaline Phosphatase Regulates Purinergic Transmission in the Central Nervous System During Development and Disease. *Comput Struct Biotechnol J* **13**, 95-100 (2015).

43. Diaz-Hernandez, M. *et al.* Tissue-nonspecific alkaline phosphatase promotes the neurotoxicity effect of extracellular tau. *J Biol Chem* **285**, 32539-48 (2010).

44. Vardy, E.R., Kellett, K.A., Cocklin, S.L. & Hooper, N.M. Alkaline phosphatase is increased in both brain and plasma in Alzheimer's disease. *Neurodegener Dis* **9**, 31-7 (2012).

45. Kellett, K.A., Williams, J., Vardy, E.R., Smith, A.D. & Hooper, N.M. Plasma alkaline phosphatase is elevated in Alzheimer's disease and inversely correlates with cognitive function. *Int J Mol Epidemiol Genet* **2**, 114-21 (2011).

46. Searles Quick, V.B., Davis, J.M., Olincy, A. & Sikela, J.M. DUF1220 copy number is associated with schizophrenia risk and severity: implications for understanding autism and schizophrenia as related diseases. *Transl Psychiatry* **5**, e697 (2015).

47. Hsu, S.C. *et al.* Mutations in SLC20A2 are a major cause of familial idiopathic basal ganglia calcification. *Neurogenetics* **14**, 11-22 (2013).

48. Taglia, I., Bonifati, V., Mignarri, A., Dotti, M.T. & Federico, A. Primary familial brain calcification: update on molecular genetics. *Neurol Sci* **36**, 787-94 (2015).

49. Figueiro-Silva, J. *et al.* Neuronal pentraxin 1 negatively regulates excitatory synapse density and synaptic plasticity. *J Neurosci* **35**, 5504-21 (2015).

50. Abad, M.A., Enguita, M., DeGregorio-Rocasolano, N., Ferrer, I. & Trullas, R. Neuronal pentraxin 1 contributes to the neuronal damage evoked by amyloid-beta and is overexpressed in dystrophic neurites in Alzheimer's brain. *J Neurosci* **26**, 12735-47 (2006).

51. Tobaben, S., Varoqueaux, F., Brose, N., Stahl, B. & Meyer, G. A brain-specific isoform of small glutamine-rich tetratricopeptide repeat-containing protein binds to Hsc70 and the cysteine string protein. *J Biol Chem* **278**, 38376-83 (2003).

52. Fonte, V. *et al.* Interaction of intracellular beta amyloid peptide with chaperone proteins. *Proc Natl Acad Sci U S A* **99**, 9439-44 (2002).

53. Mao, C.X. *et al.* Microtubule-severing protein Katanin regulates neuromuscular junction development and dendritic elaboration in Drosophila. *Development* **141**, 1064-74 (2014).

54. Yu, W. *et al.* The microtubule-severing proteins spastin and katanin participate differently in the formation of axonal branches. *Mol Biol Cell* **19**, 1485-98 (2008).

55. Zhu, J., Shang, Y. & Zhang, M. Mechanistic basis of MAGUK-organized complexes in synaptic development and signalling. *Nat Rev Neurosci* **17**, 209-23 (2016).

56. Ingason, A. *et al.* Expression analysis in a rat psychosis model identifies novel candidate genes validated in a large case-control sample of schizophrenia. *Transl Psychiatry* **5**, e656 (2015).

57. Nithianantharajah, J. *et al.* Synaptic scaffold evolution generated components of vertebrate cognitive complexity. *Nat Neurosci* **16**, 16-24 (2013).

58. Nalls, M.A. *et al.* Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. *Nat Genet* **46**, 989-93 (2014).

59. Guan, J.J. *et al.* DRAM1 regulates apoptosis through increasing protein levels and lysosomal localization of BAX. *Cell Death Dis* **6**, e1624 (2015).

60. Yu, M., Jiang, Y., Feng, Q., Ouyang, Y. & Gan, J. DRAM1 protects neuroblastoma cells from oxygen-glucose deprivation/reperfusion-induced injury via autophagy. *Int J Mol Sci* **15**, 19253-64 (2014).

61. Scarpa, J.R. *et al.* Systems Genetic Analyses Highlight a TGFbeta-FOXO3 Dependent Striatal Astrocyte Network Conserved across Species and Associated with Stress, Sleep, and Huntington's Disease. *PLoS Genet* **12**, e1006137 (2016).

62. Donlon, T.A. *et al.* FOXO3 longevity interactome on chromosome 6. *Aging Cell* (2017).

63. Sears, J.C. & Broihier, H.T. FoxO regulates microtubule dynamics and polarity to promote dendrite branching in Drosophila sensory neurons. *Dev Biol* **418**, 40-54 (2016).

64. Peng, K. *et al.* Knockdown of FoxO3a induces increased neuronal apoptosis during embryonic development in zebrafish. *Neurosci Lett* **484**, 98-103 (2010).

65. Santama, N., Er, C.P., Ong, L.L. & Yu, H. Distribution and functions of kinectin isoforms. *J Cell Sci* **117**, 4537-49 (2004).

66. Liu, X.A., Rizzo, V. & Puthanveettil, S.V. Pathologies of Axonal Transport in Neurodegenerative Diseases. *Transl Neurosci* **3**, 355-372 (2012).

67. Consortium, E. *et al.* Genome-wide association analysis of genetic generalized epilepsies implicates susceptibility loci at 1q43, 2p16.1, 2q22.3 and 17q21.32. *Hum Mol Genet* **21**, 5359-72 (2012).

68. Martins-de-Souza, D. *et al.* Proteomic analysis identifies dysfunction in cellular transport, energy, and protein metabolism in different brain regions of atypical frontotemporal lobar degeneration. *J Proteome Res* **11**, 2533-43 (2012).

69. Shulman, J.M. *et al.* Functional screening in Drosophila identifies Alzheimer's disease susceptibility genes and implicates Tau-mediated mechanisms. *Hum Mol Genet* **23**, 870-7 (2014).

70. Friede, R.L. & Samorajski, T. Axon caliber related to neurofilaments and microtubules in sciatic nerve fibers of rats and mice. *Anat Rec* **167**, 379-87 (1970).

71. Yuan, A., Rao, M.V., Veeranna & Nixon, R.A. Neurofilaments and Neurofilament Proteins in Health and Disease. *Cold Spring Harb Perspect Biol* **9**(2017).

72. Bis, J.C. *et al.* Whole exome sequencing study identifies novel rare and common Alzheimer's-Associated variants involved in immune response and transcriptional regulation. *Mol Psychiatry* (2018).

73. Marioni, R.E. *et al.* GWAS on family history of Alzheimer's disease. *Transl Psychiatry* **8**, 99 (2018).

Figure 1. Heritability and Manhattan plot of genetic variants associated with subcortical brain volumes in the European sample.

a. Family-based heritability (h^2) estimates were performed with SOLAR in the Framingham Heart Study (n = 895) and the Austrian Stroke Prevention-Family Study (n = 370). **b.** Combined Manhattan plot highlighting the most significant SNPs across all subcortical structures (nucleus accumbens = 32,562; amygdala = 34,431; brainstem = 28,809; caudate = 37,741; pallidum = 34,413; putamen = 37,571; thalamus = 34,464). Variants are colored differently for each structure (see legend in a). Linear regression models were adjusted for sex, age, age^2, total intracranial volume (CHARGE) or total brain volume (UKBB), and population stratification. The solid horizontal line denotes genome-wide significance as set in this study after additional Bonferroni correction for six independent traits ($P < 5 \times 10^{-8}/6 = 8.3 \times 10^{-9}$ for two-sided tests), the dashed horizontal line denotes the classic genome-wide threshold of $P < 5 \times 10^{-8}$. Individual Manhattan plots can be found in the Supplementary note.

Figure 2. **Partitioning heritability by functional annotation categories.**

Analyses performed in the European sample (nucleus accumbens = 32,562; amygdala = 34,431; brainstem =28,809; caudate = 37,741; pallidum = 34,413; putamen = 37,571; thalamus = 34,464). Plotted ellipses represent enrichment (proportion of h^2_g explained / proportion of SNPs in a given functional category) for subcortical structures (y-axis) across 28 functional categories (x-axis). The color bar indicates the magnitude and direction of enrichment. Starred pairs denote significant over-representation after Bonferroni correction for 168 tests (28 annotation categories and 6 independent traits, $P < 3 \times 10^{-4}$). DHS, DNase I hypersensitivity site; TSS, transcription start site.

Figure 3. **Protein-protein interaction network of 158 genes enriched for common variants influencing the volume of subcortical structures.**

The edges represent protein-protein associations, where the edge color indicates the predicted mode of action (bright green, activation; pink, posttranslational modification; red, inhibition; dark blue, binding, purple, catalysis; light blue, phenotype; black, reaction; yellow, transcriptional regulation) and the edge shape the predicted action effects (arrow, positive, flat arrow, negative; oval arrow, unspecified). Colored nodes represent the queried proteins and first shell of interactors (5 maximum), whereas white nodes represent the second shell of interactors (5 maximum).

545 **Table 1**. Genome-wide association[a] results for subcortical brain volumes in Europeans from CHARGE, ENIGMA, and the UKBB

SNP	Chr	Position	Function	A1/A2	A1 Freq.	Weight	Z-score	P[b]	Direction	I^2
Nucleus accumbens (n=32,562)										
rs9818981[c]	3	190602087	intergenic	A/G	0.09	32,282	-6.23	4.70E-10	---	63.2
rs13107325	4	103188709	missense	T/C	0.06	32,283	6.15	7.74E-10	+++	76.2
rs11747514[c]	5	65839259	intronic	T/G	0.22	32,562	-5.99	2.11E-09	---	0.0
rs868202[c]	14	56195762	intergenic	T/C	0.56	32,562	5.90	3.55E-09	+++	0.0
Amygdala (n=34,431)										
rs11111293[c]	12	102921296	intergenic	T/C	0.78	34,313	6.25	4.16E-10	+++	0.0
Brainstem (n=28,809)										
rs11111090	12	102326461	intergenic	A/C	0.52	28,809	10.79	3.70E-27	+++	0.0
rs10217651[c]	9	118923652	intronic	A/G	0.39	28,809	9.78	1.40E-22	+++	0.0
rs869640[c]	5	65015128	intronic	A/C	0.72	28,809	-8.40	4.36E-17	---	9.5
rs9398173[c]	6	109000316	intronic	T/C	0.33	28,809	-7.95	1.80E-15	---	19.0
rs10792032[c]	11	68984602	intergenic	A/G	0.49	28,648	7.75	9.08E-15	+++	39.4
rs4396983[c]	4	15132604	intergenic	A/G	0.44	28,809	-7.02	2.27E-12	---	73.6
rs9322194[c]	6	149920249	intronic	T/C	0.34	28,156	6.91	4.94E-12	+++	0.0
rs7972561[c]	12	107139983	intronic	A/T	0.33	28,809	6.90	5.05E-12	+++	0.0

rs2206656[c]	20	49130119	intronic	C/G	0.61	28,809	6.83	8.26E-12	+++	0.0
rs12479469[c]	20	61145196	intergenic	A/G	0.33	25,822	-6.80	1.08E-11	---	65.6
rs4784256[c]	16	52814559	intergenic	A/G	0.40	28,809	6.76	1.41E-11	+++	0.0
rs555925[c]	3	193544359	intergenic	T/G	0.41	27,934	6.37	1.88E-10	+++	62.9
rs12313279[c]	12	102846504	intronic	A/G	0.29	28,809	6.21	5.39E-10	+++	24.9
rs9505301[c]	6	7887131	intronic	A/G	0.89	28,691	-6.05	1.41E-09	---	43.2
rs11684404[c]	2	88924622	intronic	T/C	0.66	28,809	-5.95	2.73E-09	---	0.0
rs112178027[c]	17	27564013	intergenic	T/C	0.17	28,809	-5.90	3.67E-09	---	0.0
Caudate nucleus (n=37,741)										
rs3133370	11	92026446	intergenic	T/C	0.67	37,741	7.52	5.59E-14	+++	44.9
rs6060983[c]	20	30420924	intronic	T/C	0.70	37,741	7.04	1.95E-12	+++	0.0
rs7040561[c]	9	128528978	intronic	A/T	0.85	34,049	-6.26	3.84E-10	---	0.0
rs2817145[c]	1	3133422	intronic	A/T	0.19	35,598	6.20	5.71E-10	+++	65.3
rs148470213[c]	14	56193700	intergenic	T/C	0.54	29,429	6.18	6.48E-10	++?	0.0
rs1987471[c]	16	28825866	intergenic	T/G	0.63	37,741	5.87	4.40E-09	+++	0.0
rs12445022[c]	16	87575332	intergenic	A/G	0.33	37,741	5.87	4.45E-09	+++	0.0
rs55989340[c]	14	100635222	intergenic	A/G	0.74	37,741	-5.86	4.62E-09	---	52.0
rs4888010[c]	16	73895046	intergenic	A/G	0.47	37,741	5.86	4.67E-09	+++	74.9

rs35305377[c]	7	99938955	intronic	A/G	0.55	33,429	-5.84	5.36E-09	---	47.8
Globus pallidus (n=34,413)										
rs2923447	8	42439848	intergenic	T/G	0.59	34,413	8.11	4.88E-16	+++	34.0
rs10129414[c]	14	56193272	intergenic	A/G	0.44	34,413	-7.53	5.11E-14	---	0.0
rs196807[c]	8	24682649	intergenic	A/G	0.18	34,295	6.44	1.17E-10	+++	21.1
rs10439607[c]	20	30258541	intronic	A/G	0.30	34,413	-6.28	3.35E-10	---	0.0
rs4952211[c]	2	32611512	intronic	T/C	0.43	34,252	-5.86	4.72E-09	---	61.9
rs12567402[c]	1	21870213	intronic	T/C	0.33	34,214	5.81	6.17E-09	+++	0.0
Putamen (n=37,571)										
rs945270	14	56200473	intergenic	C/G	0.58	37,571	15.03	5.02E-51	+++	57.3
rs62098013	18	50863861	intronic	A/G	0.38	37,571	8.92	4.59E-19	+++	33.9
rs6087771	20	30306724	intronic	T/C	0.71	36,291	8.69	3.75E-18	+++	7.5
rs35200015[c]	11	117383215	intronic	A/G	0.19	37,571	-8.19	2.51E-16	---	0.0
rs1432054	11	83260225	intronic	A/G	0.64	37,571	-7.94	2.10E-15	---	0.0
rs7902527[c]	10	118715399	intronic	A/G	0.24	37,108	6.29	3.13E-10	+++	0.0
rs2244479[c]	7	50738987	intronic	T/C	0.65	36,291	-5.92	3.17E-09	---	32.1
rs2410767[c]	5	87705268	intronic	C/G	0.78	37,571	5.88	3.99E-09	+++	0.0
rs1187162[c]	11	92011126	intergenic	T/C	0.42	37,571	5.84	5.14E-09	+++	0.0

Thalamus (n=34,464)										
rs12600720[c]	17	78448640	intronic	C/G	0.69	33,023	6.25	4.06E-10	+++	0.0
rs142461330[c]	7	55012097	intergenic	T/C	0.92	34,185	-5.90	3.69E-09	---	0.0

[a] Linear regression models are adjusted for sex, age, age^2, total intracranial volume (CHARGE) or total brain volume (UKBB), and population stratification.

[b] P-values are two-tailed. Significance was set at $P < 8.3 \times 10^{-9}$ after additional Bonferroni correction for six independent traits ($5 \times 10^{-8}/6$).

[c] Novel SNPs

Chr = chromosome; Freq. = frequency of the coded allele; A1 = coded allele; A2 = non-coded allele

552 **ONLINE METHODS**

553

554 ***Study population***

555 The present effort included 53 study samples from the Cohorts of Heart and Aging
556 Research in Genomic Epidemiology (CHARGE) consortium [74], the Enhancing Neuro Imaging
557 Genetics through Meta-Analysis (ENIGMA) consortium[75], and the United Kingdom Biobank
558 (UKBB)[76]. Briefly, the CHARGE consortium is a collaboration of predominantly population-
559 based cohort studies investigating the genomics of age-related complex diseases, including
560 those of the brain (depts.washington.edu/chargeco/wiki/). The ENIGMA consortium brings
561 together various studies, approximately 75% of which are population-based, with the
562 remainder using case-control designs for various neuropsychiatric or neurodegenerative
563 diseases (enigma.ini.usc.edu/). The UKBB is a large-scale prospective epidemiological
564 study of over 500,000 individuals aged 40-69 years from the United Kingdom, established
565 to investigate the genetic and non-genetic determinants of middle and old age diseases
566 (www.ukbiobank.ac.uk/).

567 Our sample consisted of up to n=37,741 individuals of European ancestry. We
568 additionally included three generalization samples of African-Americans (up to n=769),
569 and two generalization samples of Asians (n=341). All participants have provided written
570 informed consent and participating studies obtained approval from their institutional
571 review board or equivalent organization. The institutional review boards of Boston
572 University and the University of Southern California, as well as the local ethics board of
573 Erasmus University Medical Center approved this study.

Exclusion criteria comprised prevalent dementia or stroke at the time of the MRI scan, and when available, presence of large brain infarcts or other neurological pathologies seen at the MRI that could substantially influence the measurement of brain volumes (e.g. brain tumor, trauma). Individual studies applied the exclusion criteria prior to analyses.

Definition of phenotypes

Our study investigated the volumes of seven subcortical structures: nucleus accumbens, amygdala, brainstem, caudate nucleus, globus pallidus, putamen, and thalamus. These phenotypes were defined as the mean volume (in cm^3) of the left and right hemispheres, with the exception if the brainstem that was simply defined as total volume (in cm^3). Each study contributed magnetic resonance imaging (MRI) data obtained using diverse scanners, field strengths, and acquisition protocols. The estimation of volumes for the seven subcortical brain structures and total intracranial volume was generated by freely available and in-house segmentation methods previously described and validated. Summary statistics for subcortical brain volumes in CHARGE study samples are presented in Supplementary Table 3, and the study-specific MRI protocols and software are described in Supplementary Table 5. We have recently published results describing the genetic variation associated with hippocampal volumes[20], and therefore, we have not included that brain structure in this report.

Genotyping

Genotyping was performed using a variety of commercial arrays across the participating studies. Study samples and genetic variants underwent similar quality control

597 procedures based on genetic homogeneity, call rate, minor allele frequency (MAF), and
598 Hardy-Weinberg Equilibrium. Good quality variants were used as input for imputation to
599 the 1000 Genomes Project (phase 1, version 3) reference panel[16], or the Haplotype
600 Reference Consortium (HRC, version 1.1)[17] in the UKBB, using validated software packages.
601 A detailed description of the genotyping and quality control carried by each study is
602 described in Supplementary Table 6.
603
604 ***Heritability***
605 Heritability of subcortical brain volumes was estimated in the Framingham Heart Study
606 (FHS)[77] and the Austrian Stroke Prevention Study Family Study (ASPS Fam)[78], two
607 population-based cohorts with family structure. We used SOLAR[79] to determine the ratio of
608 the genetic variance to the phenotypic variance, including variance component models that
609 were adjusted for age, sex, total intracranial volume, as well as age squared and principal
610 components if required, in the same way it is described for the genome-wide association
611 (GWA) analysis. We also estimated the variance of subcortical structures explained by SNPs
612 in a sample of n=3,486 unrelated participants from the Rotterdam Study using GCTA[80], and
613 additionally in the full European sample using LDSC regression methods[81]. Supplementary
614 Table 4 provides family- and SNP-based heritabilities for subcortical structures.
615
616 ***Genome-wide associations and meta-analysis***
617 In CHARGE and ENIGMA, each study undertook a GWA analysis on the volumes of seven
618 MRI subcortical brain structures (or those that were available to each study) according to a
619 common predefined analysis plan. Studies including unrelated participants performed

linear regression analyses, whereas those including related participants conducted linear mixed models to account for familial relationships. Models assumed additive genetic effects and were adjusted for age, sex, total intracranial volume and, if applicable, they were additionally adjusted for age^2, principal components to account for population stratification, psychiatric diagnosis (ENIGMA cohorts), and study site. Individual studies shared summary statistics to a centralized, secured computing space. Analysis in the UKBB sample followed a similar approach in n=8,312 unrelated participants although the genetic data used for these analyses uses only those variants imputed using the HRC[17] reference panel. As the data released by the UKBB did not include total intracranial volume, linear regression models in this sample are adjusted for age, age^2, sex, *total brain volume*, and principal components. We used LDSC methods[81] to investigate the genetic correlations for all subcortical structures between the CHARGE-ENIGMA and the UKBB. There was no evidence suggesting differences in the genetic architecture of both samples.

Prior to meta-analysis, we performed quality control at the study-level summary statistics using a series of quality checks implemented in EasyQC[82]. Filters were set to remove SNPs with poor imputation ($R^2 < 0.5$), rare (MAF < 0.1%), or with an effective allele count (2 x MAF x study sample size x imputation quality) < 20. Finally, we only considered variants present in at least 70% of the total European sample for each structure.

Fixed-effects meta-analyses weighting for sample size were performed using METAL[18], given that not all samples used the same methods for acquisition and post-processing of brain images. We used the LD score regression intercept to correct for population stratification and cryptic relatedness[81]. Quantile and Manhattan plots are presented for each subcortical structure in Supplementary Figure 1. To correct for multiple comparisons

643 across our seven traits, we calculated the Pearson's correlation among subcortical
644 structures adjusting for age, sex and intracranial volume in n=4,459 participants from the
645 Rotterdam Study. After 1,000 permutations, the resulting number of independent traits
646 was of six, leading to the definition of a significant threshold as $P < (5 \times 10^{-8}/6) = 8.3 \times 10^{-9}$.
647 To select our top independent SNPs in the European meta-analysis, we ran a multi-SNP-
648 based conditional & joint association analysis (GCTA-COJO)[80] using n=6,921 participants
649 from the Rotterdam Study as the reference sample. In secondary analyses, we looked for
650 the association of our index SNPs (the most significant variant in each locus) with the other
651 six subcortical structures.
652 We conducted separate meta-analyses by ancestry, and further performed a combined
653 meta-analysis including all samples. Forest plots were created to explore the contribution
654 of participating studies to each of the significant SNPs (Supplementary Figure 4). To assess
655 signal overlap with African-American and Asian samples, we first clumped variants with P
656 $< 1 \times 10^{-4}$ in the European sample, and then ran binomial sign tests for the correlation of
657 the direction of association across ethnic groups.
658
659 ***Functional annotations***
660 We used Locus Zoom[19] based on the hg 19 UCSC Genome Browser assembly for the
661 visualization of the nearest genes within a ±500 Kb genomic region. We also investigated
662 *cis* (1 Mb) expression quantitative trait loci (eQTL) and methylation QTL (meQTL) for our
663 index SNPs in post-mortem brains from the Religious Order Study and the Rush Memory
664 and Aging Project (ROSMAP). In ROSMAP, the dorsolateral prefrontal cortex (DLPFC) was
665 selected for initial multi-omics data generation, as it is relevant to multiple common

neuropathologies and cognitive phenotypes in the aging population[83]. RNA was extracted from the gray matter of DLPFC, and next-generation RNA sequencing (RNA-Seq) was done on the Illumina HiSeq for samples with an RNA integrity score > 5 and a quantity threshold > 5 ug, as previously described[83,84]. We quantile-normalized the fragments per kilobase of transcript per million fragments mapped (FPKM), correcting for batch effect with Combat[84,85]. These adjusted FPKM values were used for analysis. A subset of 407 participants had quality-controlled RNA-Seq data and were included in the eQTL analysis.

DNA methylation levels from the gray matter of DLPFC were measured using the Illumina HumanMethylation450 BeadChip, and the measurements underwent QC processing as previously described (i.e. detection $p < 0.01$ for all samples)[83], yielding 708 participants with 415,848 discrete CpG dinucleotide sites with methylation measurement. Any missing methylation levels from any of quality-controlled CpG dinucleotide sites were imputed using a k-nearest neighbor algorithm for $k = 100$[83]. A subset of 488 participants in our study had quality-controlled genome-wide methylation data and were included in the *cis*-methylation QTL analysis. Finally, the associations between our index SNPs and CpG sites were plotted along Roadmap Epigenomic chromatin states for ten brain tissues[86].

We further queried *cis* and *trans* eQTLs in non-brain and brain tissues from additional eQTL repositories[87]. We searched for proxies to our index SNPs with a $r^2>0.8$ using the European population reference in rAggr (1000G, phase 1, Mar 2012), and then queried index and proxy SNPs against eQTLs from diverse databases.[88] Blood cell related eQTL studies included fresh lymphocytes and leukocytes, leukocyte samples in individuals with Celiac disease, whole blood samples, lymphoblastoid cell lines (LCL) derived from asthmatic children, HapMap LCL from 3 populations, a separate study on HapMap CEU LCL,

LCL population samples, neutrophils, CD19+ B cells, primary PHA-stimulated T cells, CD4+ T cells, peripheral blood monocytes, long non-coding RNAs in monocytes and CD14+ monocytes before and after stimulation with LPS or interferon-gamma, CD11+ dendritic cells before and after *Mycobacterium tuberculosis* infection and a separate study of dendritic cells before or after stimulation with lipopolysaccharide (LPS), influenza or interferon-beta; micro-RNA QTLs, DNase-I QTLs, histone acetylation QTLs, and ribosomal occupancy QTLs were also queried for LCL; splicing QTLs and micro-RNA QTLs were queried in whole blood. Non-blood cell tissue eQTL searches included omental and subcutaneous adipose, visceral fat stomach, endometrial carcinomas, ER+ and ER- breast cancer tumor cells, liver, osteoblasts, intestine and normal and cancerous colon, skeletal muscle, breast tissue (normal and cancer), lung, skin, primary fibroblasts, sputum, pancreatic islet cells, prostate, rectal mucosa, arterial wall and heart tissue from left ventricles and left and right atria. Micro-RNA QTLs were also queried for gluteal and abdominal adipose and liver. Methylation QTLs were queried in pancreatic islet cells. Further mRNA and micro-RNA QTLs were queried from ER+ invasive breast cancer samples, colon-, kidney renal clear-, lung- and prostate-adenocarcinoma samples. Brain eQTL studies included brain cortex, cerebellar cortex, cerebellum, frontal cortex, gliomas, hippocampus, inferior olivary nucleus (from medulla), intralobular white matter, occiptal cortex, parietal lobe, pons, pre-frontal cortex, putamen (at the level of anterior commussure), substantia nigra, temporal cortex, thalamus and visual cortex. eQTL data was integrated from online sources including ScanDB[89], the GTEx Portal[90], and the Pritchard Lab[91]. Cerebellum, parietal lobe and liver eQTL data was downloaded from ScanDB and cis-eQTL were limited to those with $P<1.0 \times 10^{-6}$ and trans-eQTLs with $P < 5.0$

712 × 10^{-8}. Results for GTEx Analysis V6 for 48 tissues were downloaded from the GTEx Portal
713 (www.gtexportal.org). For all gene-level eQTL, if at least 1 SNP passed the tissue-specific
714 empirical threshold in GTEx, the best SNP for that eQTL was always retained.
715
716 ***Associations of cognition and neuropathology phenotypes with gene expression in***
717 ***brain***
718 We further related cognitive function and neuropathological findings to the expression
719 of the 199 gene set influencing subcortical volumes in 508 brains from the ROSMAP
720 samples.
721 Briefly, brain autopsies were performed as previously described and each brain was
722 inspected for common pathologies relating to loss of cognition in aging populations[92,93]. In
723 this report, we included: neurofibrillary tangles, neuritic plaques, β-amyloid load, tau
724 density, hippocampal sclerosis, Lewy bodies and neuronal loss in substantia nigra.
725 Neurofibrillary tangles and neuritic plaques were visualized by modified Bielschowsky
726 silver stain, then counted and scaled in five brain regions: mid-frontal, temporal, inferior
727 parietal, entorhinal cortex, and hippocampus CA1. Composite scores for each of these three
728 pathology types were derived by scaling the counts within each of the five regions, and
729 taking the square root of the average of the regional scaled values to account for their
730 positively skewed distribution[92-94]. β-amyloid load and tau tangle density were measured
731 by immunohistochemistry and square root transformed as previously described[95]. Lewy
732 bodies were identified using immunohistochemistry and were further dichotomized as
733 present or absent based on the recommendations of the Report of the Consortium on DLB
734 International Workshop[96]. Hippocampal sclerosis was recorded as either present or absent

as evaluated with H&E stain. Nigral neuronal loss was assessed in the substantia nigra in the mid to rostral midbrain near or at the exit of the 3rd nerve using H&E stain and 6 micron sections using a semi-quantitative scale (0–3)[97].

Global cognition was computed as a composite score of 19 (ROS) and 17 (MAP) cognitive tests performed at annual evaluations including five cognitive domains: episodic memory, semantic memory, working memory, perceptual speed, and visuospatial ability[92,93]. From these scores, we created normalized summary measures to limit the influence of outliers. We used global cognition proximate to death to derive cognitive reserve. Separately, the residual slope of global cognitive change and the residual slopes of cognitive change in the five cognitive domains were derived through general linear mixed models, controlling for age at enrollment, sex, and education.

Phenotypic and genetic correlations

We estimated the Pearson's partial phenotypic correlations among the volumes of subcortical structures in 894 participants from the Framingham Heart Study. Similarly, to the GWA, these analyses were corrected for the effects of sex, age, age^2, total intracranial volume and PC1.

Genetic correlation analyses were performed using LDSC regression methods[81]. The GWA meta-analysis results for the seven subcortical brain structures were correlated with each other's, as well as with published GWA studies on the following traits: hippocampal volume[20], intracranial volume[21], white matter hyperintensities[22], stroke subtypes[23], adult height and body mass index[24], fat-free mass and whole-body water mass[98], Alzheimer's

757 disease[26], Parkinson's Disease[27], general cognitive function[25], bipolar disorder and
758 schizophrenia[28], and ADHD[29].
759
760 ***Look-up of functional orthologs in Drosophila melanogaster***
761 For the cross-species assessment of gene-phenotype relationships in *Drosophila*, we
762 relied on a similar analytic approach as in prior work[99]. Human genes were mapped to
763 corresponding *Drosophila* orthologs using DIOPT: Drosophila Integrated Ortholog
764 Prediction Tool (www.flyrnai.org/diopt)[100], which incorporates 14 distinct algorithms to
765 define orthology. Fly gene orthologs were defined based on a DIOPT score of 2 or greater,
766 indicating at least 2 algorithms were in agreement on the pairing. When more than one of
767 the fly ortholog was predicted, all such genes meeting this threshold were included in our
768 analyses. This resulted in a gene set consisting of 168 *Drosophila* homologs of human
769 candidate genes at subcortical volume susceptibility loci. The resulting 37 genes associated
770 with "neuroanatomy defective" phenotypes in *Drosophila* (22%) were annotated based on
771 the controlled vocabulary terms implemented in FlyBase (flybase.org/)[101]. Genes causing
772 "neuroanatomy defective" phenotypes in *Drosophila* include both loss- or gain-of-function
773 genetic manipulations of fly gene homologs. Loss-of-function studies included both
774 classical mutant alleles (e.g. point mutations, gene deletions, or transposon insertions) or
775 gene knockdown using RNA interference transgenic strains. Gain-of-function experiments
776 were based on tissue specific overexpression of the fly gene orthologs. The hypergeometric
777 overlap test was used to assess for enrichment of "neuroanatomy defective" phenotypes
778 among the conserved gene set.
779

780 ***Protein-protein interactions and network analysis***

781 We used the human STRING database resource (string-db.org)[32] for the exploration of
782 direct (physical) and indirect (functional) protein-protein interactions based on the gene
783 set derived from the GWA results and functional annotations (Supplementary Table 13).
784 The input parameters included a medium-confidence interaction scores (0.4) with first and
785 second shells of maximum 5 interactors. Finally, we generated a protein-protein
786 interaction network based on known and predicted interactions.

787

788 ***Partitioning heritability***

789 Partitioned heritability was estimated with stratified LDSC methods[30]. This method
790 partitions SNP heritability using GWAS summary results and accounting by LD. We used
791 the meta-analysis results from the European sample to partitioning SNPs by 28 functional
792 categories, including: coding, intron, promoter, 3'/5' UTRs, digital genomic footprint (DGF),
793 transcription factor binding sites, chromHMM and Segway annotations for six cell lines,
794 DNase I hypersensitivity sites (DHS), H3K4me1, H3K4me3 and H3K9ac marks, two sets of
795 H3K27ac marks, super-enhancers, conserved regions in mammals, and FANTOM5
796 enhancers. Significance was set at $P < (0.05/(28 \times 6)) = 3 \times 10^{-4}$.

797

798 ***Data availability***

799 The genome-wide summary statistics that support the findings of this study will be made
800 available through the CHARGE dbGaP (accession number phs000930) and ENIGMA
801 (http://enigma.ini.usc.edu/research/download-enigma-gwas-results) websites.

802

METHODS REFERENCES

74. Psaty, B.M. *et al.* Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Consortium: Design of prospective meta-analyses of genome-wide association studies from 5 cohorts. *Circ Cardiovasc Genet* **2**, 73-80 (2009).

75. Thompson, P.M. *et al.* The ENIGMA Consortium: large-scale collaborative analyses of neuroimaging and genetic data. *Brain Imaging Behav* **8**, 153-82 (2014).

76. Sudlow, C. *et al.* UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PLoS Med* **12**, e1001779 (2015).

77. Tsao, C.W. & Vasan, R.S. Cohort Profile: The Framingham Heart Study (FHS): overview of milestones in cardiovascular epidemiology. *Int J Epidemiol* **44**, 1800-13 (2015).

78. Schmidt, R. *et al.* Assessment of cerebrovascular risk profiles in healthy persons: definition of research goals and the Austrian Stroke Prevention Study (ASPS). *Neuroepidemiology* **13**, 308-13 (1994).

79. Almasy, L. & Blangero, J. Multipoint quantitative-trait linkage analysis in general pedigrees. *Am J Hum Genet* **62**, 1198-211 (1998).

80. Yang, J., Lee, S.H., Goddard, M.E. & Visscher, P.M. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet* **88**, 76-82 (2011).

81. Bulik-Sullivan, B.K. *et al.* LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet* **47**, 291-5 (2015).

82. Winkler, T.W. *et al.* Quality control and conduct of genome-wide association meta-analyses. *Nat Protoc* **9**, 1192-212 (2014).

83. Bennett, D.A., Yu, L. & De Jager, P.L. Building a pipeline to discover and validate novel therapeutic targets and lead compounds for Alzheimer's disease. *Biochem Pharmacol* **88**, 617-30 (2014).

84. Chan, G. *et al.* CD33 modulates TREM2: convergence of Alzheimer loci. *Nat Neurosci* **18**, 1556-8 (2015).

85. Johnson, W.E., Li, C. & Rabinovic, A. Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics* **8**, 118-27 (2007).

86. Roadmap Epigenomics, C. *et al.* Integrative analysis of 111 reference human epigenomes. *Nature* **518**, 317-30 (2015).

87. Eicher, J.D. *et al.* GRASP v2.0: an update on the Genome-Wide Repository of Associations between SNPs and phenotypes. *Nucleic Acids Res* **43**, D799-804 (2015).

88. Zhang, X. *et al.* Synthesis of 53 tissue and cell line expression QTL datasets reveals master eQTLs. *BMC Genomics* **15**, 532 (2014).

89. Zhang, W. *et al.* SCAN database: facilitating integrative analyses of cytosine modification and expression QTL. *Database (Oxford)* **2015**(2015).

90. Consortium, G.T. The Genotype-Tissue Expression (GTEx) project. *Nat Genet* **45**, 580-5 (2013).

91. Veyrieras, J.B. *et al.* High-resolution mapping of expression-QTLs yields insight into human gene regulation. *PLoS Genet* **4**, e1000214 (2008).

92. Bennett, D.A. *et al.* Overview and findings from the rush Memory and Aging Project. *Curr Alzheimer Res* **9**, 646-63 (2012).

93. Bennett, D.A., Schneider, J.A., Arvanitakis, Z. & Wilson, R.S. Overview and findings from the religious orders study. *Curr Alzheimer Res* **9**, 628-45 (2012).

94. Replogle, J.M. *et al.* A TREM1 variant alters the accumulation of Alzheimer-related amyloid pathology. *Ann Neurol* **77**, 469-77 (2015).

95. Barnes, L.L., Schneider, J.A., Boyle, P.A., Bienias, J.L. & Bennett, D.A. Memory complaints are related to Alzheimer disease pathology in older persons. *Neurology* **67**, 1581-5 (2006).

96. McKeith, I.G. *et al.* Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB): report of the consortium on DLB international workshop. *Neurology* **47**, 1113-24 (1996).

97. Schneider, J.A. *et al.* Substantia nigra tangles are related to gait impairment in older persons. *Ann Neurol* **59**, 166-73 (2006).

98. Zheng, J. *et al.* LD Hub: a centralized database and web interface to perform LD score regression that maximizes the potential of summary level GWAS data for SNP heritability and genetic correlation analysis. *Bioinformatics* **33**, 272-279 (2017).

99. Wangler, M.F., Hu, Y. & Shulman, J.M. Drosophila and genome-wide association studies: a review and resource for the functional dissection of human complex traits. *Dis Model Mech* **10**, 77-88 (2017).

100. Hu, Y. *et al.* An integrative approach to ortholog prediction for disease-focused and other functional studies. *BMC Bioinformatics* **12**, 357 (2011).

101. Marygold, S.J., Crosby, M.A., Goodman, J.L. & FlyBase, C. Using FlyBase, a Database of Drosophila Genes and Genomes. *Methods Mol Biol* **1478**, 1-31 (2016).

Editorial summary:

Genome-wide analysis identifies variants associated with the volume of seven different sub-cortical brain regions defined by magnetic resonance imaging. Implicated genes are involved in neurodevelopmental and synaptic signaling pathways.